

Abstract

Phytochemicals are increasingly being used in cancer treatment due to affordability and potential anticancer effect with minimal adverse reactions compared to chemotherapy. *Markhamia tomentosa* is a medicinal plant used traditionally to treat cancer. In this study, the antiproliferative compounds from *M. tomentosa* were isolated using bioactivity-guided approach and characterized using various spectroscopic techniques. Through bioassay-guided fractionation of the crude ethanolic leaf extract, the dichloromethane (Mdf) and ethyl-acetate (Mef) fractions exhibited potent cytotoxicity activity against HeLa cells with IC₅₀ values of 83.26 and 104.5 µg/ml respectively in the MTT assay. Trypan blue assay and cell cycle analysis showed that Mdf fraction demonstrated cytotoxic effect with more extensive cell death and induced G₀/G₁ phase cell cycle arrest with concomitant decrease in S phase. Mef fraction showed reduced percentage of stained dead cells as compared to Mdf fraction and induced G₂/M phase with increase in the size of sub-G₁ phase, corresponding to apoptosis. From the isolation and purification of Mdf and Mef fractions by repeated column chromatography, followed by identification by 1D and 2D NMR spectroscopy and by comparison of the NMR data with values reported in literature, sitosterol 1, mollic acid 2, phytol 3 and oleanolic acid 4 were isolated for the first time from *M. tomentosa*. Mollic acid 2 exhibited more potent cytotoxic activity compared to compounds 1, 3 and 4. The results from these findings suggest that mollic acid 2 isolated from Mef which exhibited apoptotic cell death may be responsible for the earlier reported apoptosis induction capability of *M. tomentosa* against cervical cancer cell line HeLa.