

HAEMATOLOGICAL AND SERUM ENZYMES BIOMARKERS OF HEAVY METALS IN *Chrysichthys nigrodigitatus* AND *Cynoglossus senegalensis*

Ayoola, S. O and Adekunbi, F. O

Department of Marine Sciences, University of Lagos, Lagos State, Nigeria  
Corresponding Author: Email: sayoola@unilag.edu.ng, soayoola@yahoo.com,  
Tel: 234(80)34650102

ABSTRACT

Heavy metals are constituents of marine environment that occur as a result of pollution principally due to the discharge of untreated wastes into water bodies by many industries. Haematological, serum enzymes activity and heavy metal concentrations in feral populations of *Chrysichthys nigrodigitatus* (Lacepede:1803) and *Cynoglossus senegalensis* (Kaup: 1858) from Ebute-Oba section of the Lagos Lagoon, were evaluated from July-September, 2012 in order to determine and establish the relationship between heavy metal levels, serum enzymes activity and haematological parameters in *Chrysichthys nigrodigitatus* and *Cynoglossus senegalensis*. Levels of metals in *C. nigrodigitatus* and *C. senegalensis* in mg/l were as follows: (Cr:  $0.07 \pm 0.001$ , Cu:  $0.027 \pm 0.01$ , Fe:  $0.481 \pm 0.214$ , Pb:  $0.001 \pm 0.001$ , Zn:  $0.316 \pm 0.575$ ) and (Cr:  $0.010 \pm 0.07$ , Cu:  $0.029 \pm 0.01$ , Fe:  $0.467 \pm 0.217$ , Pb:  $0.021 \pm 0.044$ , Zn:  $0.433 \pm 0.499$ ) respectively. Serum alanine aminotransferase (ALT), Aspartate aminotransferase (AST) and Alkalinephosphase (ALP) activities in *C. nigrodigitatus* and *C. senegalensis* measured in IU/l were  $64.995 \pm 44.123$ ,  $16.345 \pm 166$ ,  $157.787 \pm 34.88$  and  $73.758 \pm 48.768$ ,  $29.532 \pm 20.100$ ,  $45.707 \pm 285$  respectively. ALP activity between the two species showed a significance difference ( $P < 0.05$ ). Haematological parameters recorded for *C. nigrodigitatus* were Hb with  $13.325 \pm 8.293$ g/dl, PCV ( $25.53 \pm 11.79\%$ ), RBC ( $2.95 \pm 0.73 \times 10^6 \text{mm}^3$ ), WBC ( $124.19 \pm 56.41 \times 10^4 \text{mm}^3$ ), MCHC ( $32.46 \pm 2.18$ g/l), Neutrophils ( $41.00 \pm 27.91\%$ ) and Lymphocyte ( $64.33 \pm 31.94\%$ ). While values for *C. senegalensis* were: Hb ( $11.33 \pm 8.04$  g/dl), PCV ( $34.00 \pm 22.83\%$ ), RBC ( $2.70 \pm 1.98 \times 10^6 \text{mm}^3$ ), WBC ( $275.20 \pm 167.27 \times 10^4 \text{mm}^3$ ), MCV ( $178.94 \pm 126$ fl), MCH ( $59.63 \pm 44.50$  Pg), MCHC ( $33.33 \pm 33.28$ g/l), Neutrophils ( $39.00 \pm 30.33\%$ ) and Lymphocytes ( $80.00 \pm 71.77\%$ ). Accumulation of heavy metals in both fish species leads to an increased ALT, AST and decreased ALP activities. Conversely, accumulation resulted in reducing Haemoglobin and an increased Red Blood Cells and Lymphocyte counts. Haematological and serum enzymes activities are predictive biomarkers for the detection and monitoring of aquatic ecosystems pollution. Therefore, there is high level of pollution in the area and is of environmental concern for aquatic organism.

**Keywords:** Biomarkers, Haematological, Serum Enzymes, Heavy Metals, Fish

INTRODUCTION

Aquatic environment are exposed to a number of pollutants that are mainly released from effluents discharged from industries, sewage treatment plants, and drainage from urban and agricultural areas. These pollutants cause serious damage to aquatic life (Karbassi *et al.*, 2006). The tremendous increase in the use of heavy metals over the past few decades has inevitably resulted in an increased flux of metallic substances in the aquatic environment (Yang and Rose, 2003).

A large part of these elements exert their toxic effects by generating reactive oxygen species (ROS), causing oxidative stress. Most of the heavy metal ions are toxic or carcinogenic in nature and pose a threat to human health and the environment. According to Olatunji and Abimbola (2010), the city of Lagos is the most populous in Nigeria and is projected to be the fifth most populous in the world by 2015. The city harbours over 75% of the industrial outfits within the country which include textile industries, chemical and paint industries, breweries and bottling companies, metal industries, ship yards,

plastic and petro-chemical factories, paper mill and sawmills. These industries discharge untreated wastes into the several drainages and canals that find their way into the Lagos lagoon (Okoye, 1989).

The Lagos Lagoon which lies within longitudes  $6^{\circ}25'$  and  $6^{\circ}43'E$  and latitudes  $3^{\circ}22'$  and  $3^{\circ}40'N$  is the largest of the three lagoon systems occurring in the Lagos area receiving over 80% of the land-derived run-offs laden with various types of wastes. An estimated  $10,000 \text{m}^3$  of industrial effluents are discharged into the Lagos Lagoon per day (Singh *et al.*, 1995; Don-Pedro *et al.*, 2004).

However, there has been growing interests in the determination of the levels of heavy metals in fish (Kalay *et al.*, 1999). Fishes are good bioindicators of the degree of pollution in the aquatic environment due to their propensity to bio-accumulate toxic metals and other contaminants in their internal organs, most especially their edible muscle tissues which could reach toxic concentrations leading to serious health hazards.



Fish has attracted much attention in biomonitoring water pollution due to its special biological characters such as relatively big body size, long life cycle, easy to raise etc. More importantly fish species are at the top position in the aquatic food chain and may directly affect the health of humans, which make it much significant for biomonitoring. Fish which is at high trophic level of the aquatic food chain accumulate substantial amount of metals in their soft and hard tissues (Mansour and Sidky, 2002). Pollutants enter fish through a number of routes: via skin, gills, oral consumption of water, food and non-food particles. Once absorbed, pollutants are transported in blood stream to either a storage point i.e. (bone) or to the liver for transformations and/or storage (Obasohan, 2008). Heavy metals are high priority pollutants because of their relative high toxicity and persistent nature in the environment. A sound understanding of dynamics of the concentration and distribution of heavy metals and their compounds in various compartments of the environment is a priority for good environmental management programmes all over the world (Don-pedro *et al.*, 2004). Hence it is important to regularly monitor the levels of metals and other contaminants that are being discharged into the aquatic ecosystems which can in turn be accumulated by fisheries resources. This study was undertaken to examine the relationship

between heavy metal levels with alanine amino transferase (ALT), Aspartate amino transferase (AST), Alkaline phosphatase (ALP) activities and haematological parameters in *Chrysichthys nigrodigitatus* and *Cynoglossus senegalensis*.

## MATERIALS AND METHODS

### Description of study area

Ebute-Oba is a fish landing site located along the shores of the Lagos Lagoon, situated between latitudes  $6^{\circ}27'58.3''$  N and  $6^{\circ}27'55.9''$  N and longitudes  $3^{\circ}29'32.2''$  E and  $3^{\circ}23'49.0''$  E in the vicinity of the Third Mainland Bridge in Isale-Eko Lagos Island (Fig.1). The landing site experienced similar environmental conditions characteristic of the Lagos Lagoon, notably semi-diurnal tidal regime and seasonal salinity variations due to rainfall. Both finfish and shell fish commonly found in the market around Lagos Island come from landings from this site. Fisher- folk which comprise of Eguns make their catch from a wide expanse of the Lagos Lagoon making use of cast and seine nets with motorized canoes. Amongst the notable species caught and landed are; *Chrysichthys nigrodigitatus*, *Ilisha africana*, *Cynoglossus senegalensis*, *Penaeus notialis*, *Machrobrachium machrobrachium* and *M. vollehovenii*.

A common scene around this landing site is a constant vehicular movement which is predominant throughout the day.

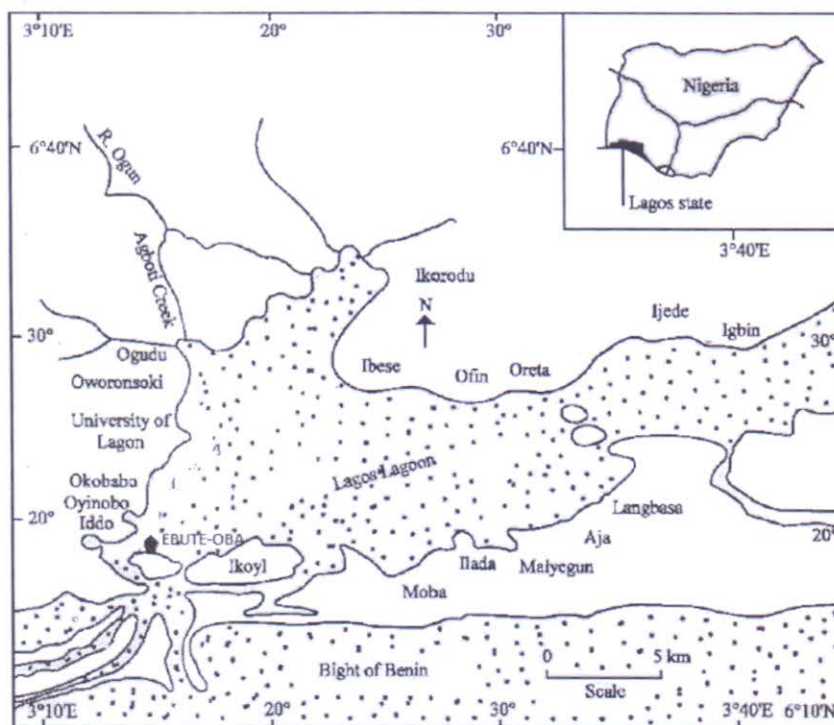


FIG 1: Map of Lagos Lagoon showing study location.



### Field sampling and measurement of morphometric features

Samples of *Chrysichthys nigrodigitatus* and *Cynoglossus senegalensis* were collected twice a month between July and September, 2012 between 09.00 – 12.00 hours on each sampling day. Total length (cm) and body weight (g) was measured using a measuring board and an electric weighing balance respectively. A total of 30 samples of *C. nigrodigitatus* and 30 samples of *C. senegalensis* were collected.

### Blood collection

Blood was collected from the caudal blood vessels on the field to avoid stress and this lasted for about 5 minutes. After blood collection, fish samples were transported to the ecotoxicology laboratory of the Marine Sciences Department, University of Lagos in ice-chest for heavy metal analysis of the tissues.

Surface water samples were collected using 1 litre water sampling bottles. The bottles were preserved in a cool dry place and transported to the laboratory for further analysis of water quality parameters not measured in-situ. Parameters measured in-situ include; surface water temperature, hydrogen ion concentration, electrical conductivity, salinity, turbidity and dissolved oxygen. Chemical oxygen demand was measured ex-situ in the laboratory. The surface water temperature of the water sample was determined in-situ by using mercury-in-glass thermometer calibrated in degree centigrade ( $^{\circ}\text{C}$ ). The thermometer was immersed in the water for about five (5) minutes and the height of the mercury was read on the graduated glass tube. The pH measures hydrogen ion activity in water. The pH of the water was determined with the aid of a pH meter. This was done by dipping the probe in the water and reading off the values from the meter. The conductivity of the water samples was measured using the Horiba U-10 water quality checker and values were read off from the meter. This was done in-situ with the use of a hand held refractometer. A drop of the water sample was placed on the prism of the meter of the refractometer and the meter was adjusted to  $\infty$  mark and viewed through the eyepiece. The daylight plate was closed and the salinity of the water sample read on the scale. Turbidity is a measure of the amount of light. This was measured at the study site using the Horiba water checker, model U-10 probe. The dissolved oxygen level was measured in-situ using a DO meter. The probe was placed in the water and the reading was recorded from the meter. The measurement of COD is based on the principle that almost all organic compounds in water can be oxidized to  $\text{CO}_2$  and water by the action of strong oxidizing agents under acid condition. This was done in the laboratory.

### Laboratory analyses of heavy metals

Determination of Heavy Metals in Fish Samples were carried out using Atomic Absorption Spectrophotometer (AAS).

Metal concentrations for all extracts were determined using Perkin Elmer Model 200 Atomic Absorption Spectrophotometer (air-acetylene flame). Accuracy was assessed by analyzing three replicates of selected samples yielding standard deviations lower than 10% for all the metals (Cd, Cr, Cu, Fe, Pb and Zn).

### Determination of heavy metals in water samples

Water sample is first aspirated into the flame or electrothermal device where it is vaporized and atomized. Radiation of the proper wavelength is then passed through the vapour containing the ground state atoms of the metal where absorption occurs. The magnitude of the AAS absorption signal is directly proportional to the concentration of the analyzed metal in the sample solution.

### Statistical analysis

Variations in heavy metal levels in *C. nigrodigitatus*, *C. senegalensis* and water were tested by one-way Analysis of Variance (ANOVA) at 5% level of significance. Post-hoc comparison of means between fish species were carried out using Least Square Difference (LSD) test to determine which values differed significantly. Comparison of mean heavy metal levels between *C. nigrodigitatus* and *C. senegalensis* was carried out by independent T-test. Pearson correlation analysis was carried out to determine the relationship between heavy metals in fish, serum enzymes activity and haematological parameters. The analysis was performed using the SPSS 17.0 package.

## RESULTS

### Heavy metals in fish species and water

Heavy metals contents in the flesh of *Chrysichthys nigrodigitatus*, *Cynoglossus senegalensis* and water during the sampling periods is presented in tables 1. The result on the table reveals that heavy metals concentration in the flesh of both fish species and water is generally low and are below WHO/FAO/FEPA allowable limits. No Cadmium was detected in all matrices measured.

Analysis of Variance (ANOVA) of the mean values of the heavy metals concentration of *C. nigrodigitatus*, *C. senegalensis* and water showed a significant difference. There was a significant difference ( $P < 0.05$ ) in Cr and Fe between *C. nigrodigitatus* and water for the sampling periods. There was a significant difference ( $P < 0.05$ ) between Fe concentration in *C. senegalensis* and water all through the sampling period. The result of serum enzymes activity in fish species is presented in table 2. T-



test analysis showed no statistical significant difference ( $P>0.05$ ) between mean serum AST activity value in *C.nigrodigitatus* ( $65.00\text{U/L}\pm44.12$ ) and *C.senegalensis* ( $73.76\text{U/L}\pm48.77$ ). No statistical difference ( $P>0.05$ ) was recorded between mean serum ALT activity value in *C. nigrodigitatus* ( $16.35\text{ IU/L}\pm12.16$ ) and *C. senegalensis* ( $29.53\text{U/L}\pm20.10$ ). A statistical significant difference was recorded ( $P< 0.05$ ) between mean serum ALP activity value in *C.nigrodigitatus* ( $157.79\text{ IU/L}\pm34.89$ ) and *C.senegalensis* ( $45.71\text{U/L}\pm35.29$ ) for the

sampling periods. Haematological parameters in fish species are presented in table 3. T-test showed no statistically significant difference ( $P>0.05$ ) for all haematological parameters recorded between *C. nigrodigitatus* and *C. senegalensis*. The Morphometric parameters are presented in table 4. Water Quality Parameters of the study area are presented in table 5. Pearson Rank Correlation ( $P=0.05$ ) showed no statistical significance ( $P>0.05$ ) for all water quality parameters recorded for the sampling periods.

**Table 1: Heavy metals concentration in flesh of *C. nigrodigitatus*, *C.senegalensis* and water**

Species	Mean $\pm$ SD ( mg/l)					
	Cd	Cr	Cu	Fe	Pb	Zn
<i>C.nigrodigitatus</i>	-	$0.007\pm0.00$	$0.027\pm0.01$	$0.481\pm0.21$	$0.001\pm0.00$	$0.316\pm0.57$
<i>C.senegalensis</i>	-	$0.010\pm0.00$	$0.029\pm0.01$	$0.467\pm 0.21$	$0.021\pm0.04$	$0.433\pm0.49$
WHO Guidelines for fish	2.0	0.15	0.15-1.0	-	2.0	1.5
Water	-	$0.014\pm0.00$	$0.020\pm 0.04$	$1.098\pm 0.44$	$0.001\pm0.00$	$0.458\pm0.68$
WHO Guidelines for Water	0.005	0.05	-	-	0.01	3.0

**Table 2: Serum enzymes activity in *C. nigrodigitatus* and *C. Senegalensis***

Species	Mean IU/L $\pm$ SD		
	AST	ALT	ALP
<i>C.nigrodigitatus</i>	$64.995\pm44.123$	$16.345\pm12.166$	$157.787\pm34.88$
<i>C.nigrodigitatus</i>	$73.758\pm48.768$	$29.532\pm20.100$	$45.707\pm35.285$

**Table 3: Haematological Parameters in *C. nigrodigitatus* and *C.senegalensis***

Species	Mean $\pm$ SD											
	Hb g/dl	PCV %	RBC $10^6\text{mm}^{-3}$	WBC $10^4\text{mm}^{-3}$	MCV fL	MCH Pg	MCHC g/l	Neutrophil %	Lymphocytes %	Monocytes %	Eosinophils %	Basophils %
<i>C.nirodigitatus</i>	$13.32\pm8.29$	$25.53\pm11.79$	$2.95\pm0.73$	$124.19\pm56.41$	$97.89\pm50.56$	$36.46\pm12.95$	$32.46\pm2.18$	$41.00\pm27.91$	$64.33\pm31.94$	-	-	-
<i>C.senegalensis</i>	$11.33\pm8.04$	$34.00\pm22.83$	$2.70\pm1.98$	$275.20\pm167.27$	$178.94\pm126$	$59.63\pm44.50$	$33.33\pm33.28$	$39.00\pm30.33$	$80.00\pm71.17$	-	-	-

**Table 4: Length and weight of *C. nigrodigitatus* and *C. senegalensis***

Species	Morphometric Parameters	Range	Minimum	Maximum	Mean of Standard Deviation
<i>C.nigrodigitatus</i>	Length (cm)	27.000	26.000	53.000	$37.41\pm9.52$
	Weight (g)	1133.000	522.000	1655.000	$859.50\pm40.9$
<i>C.senegalensis</i>	Length (cm)	14.000	29.000	43.000	$36.93\pm5.48$
	Weight (g)	942.000	473.000	1415.000	$748.71\pm35.15$

**Table 5: Water quality parameters**

Parameters	Range	Minimum	Maximum	Mean Standard Deviation
Water Temperature ( $^{\circ}\text{C}$ )	1.00	24.00	25.00	$24.17\pm0.41$
Dissolved Oxygen (mg/l)	0.57	1.40	1.97	$1.71\pm0.22$
PH	1.50	6.30	7.80	$7.08\pm0.59$
Conductivity ( $\mu\text{Scm}$ )	1100.00	6900.00	8000.00	$7650.00\pm33.70$
Turbidity( FTU)	2.00	1.00	3.00	$2.00\pm0.89$
Salinity ( ‰)	2.00	3.00	5.00	$4.17\pm0.98$
COD ( mg/l)	16.00	24.00	24.00	$32.33\pm5.43$

## DISCUSSION

Bioaccumulation of heavy metals in tissues of marine organisms has been identified as an indirect measure of the abundance and availability of metals in the marine environments (Kucuksegin *et al.*, 2006). For this reason, monitoring fish tissue contamination serves an

important function as an early warning indicator of sediments contamination or related water quality problems ( Mansour and Sidky, 2002; Babatunde, 2012) and enables appropriate action to protect public health and the environment. Several studies (Ademoroti 1996; Allen 1995; Karthikeyan *et al.*, 2007) have



indicated that fish are able to accumulate and retain heavy metals from their environment. Adeyeye et al., (1996) also showed that the concentration of metals was a function of fish species as it accumulates more in some fish species than others. Apparently, *C. nigrodigitatus* and *C. senegalensis* accumulated heavy metals in the flesh with essential metals in higher levels than non-essential metals. The average concentrations of Cd, Cr, Cu, Fe, Pb and Zn in flesh of the two species showed the following trend (*C. nigrodigitatus*); Fe>Zn>Cu>Cr>Pb>Cd and (*C. senegalensis*); Fe>Zn>Cu>Pb>Cr>Cd. Except for Fe, mean concentration of heavy metals were higher in *C. senegalensis* when compared to *C. nigrodigitatus*. From the study, the concentration of Cr, Pb, and Zn in *C. nigrodigitatus* and *C. senegalensis* showed lower levels compared to WHO maximum permissible limits. An important observation was the elevated levels of metals especially Cr, Fe and Zn in both species which could be directly attributable to bioaccumulation as evidently depicted by the accumulation factors. Metals (Copper and Zinc) play an important role in cellular metabolism acting as co-factor in a number of important enzymes. However, they can become toxic when elevated concentrations are introduced into the environment (Younis et al., 2012). The concentrations of the metals (Cd, Cr, Cu, Fe, Pb, and Zn) in flesh of the two fishes were lower than WHO limit and pose no potential threats to public health.

Pollution monitoring using enzyme inducement or enzyme dispersion in fish or other aquatic organisms has been proposed for studying polluted environments. Various organic and inorganic wastes in industrial and domestic effluents are responsible for water pollution. Conventional laboratory toxicity studies cannot be extrapolated to the natural environment because they lack natural ecological realism (Abdelmeguid et al., 2002). Since fish often respond to toxicants in a similar way to higher vertebrates they can be used to screen for chemicals that are potentially teratogenic and carcinogenic in humans. Plasma enzymes depends on the rate of release of enzymes from damaged cells, which in turn depend on the rate at which damage is occurring and at the extent of cell damage. The varied levels of aminotransferase observed in the two species investigated in the present study when compared to the mean levels recorded in other species (Younis et al., 2012; Aliakbar et al., 2010) suggest high activity levels in these species. It has been reported that the variation in enzyme activities in heavy metals exposed fish is due to increased permeability of the cells as well as the direct effect of heavy metal on the tissues (Roy, 2002).

The pattern of AST and ALT activities recorded in this study showed that cellular

damage could arise due to chronic exposure to heavy metals in the environment of the test species. No significant difference between plasma enzymes activity between the fish species occurred in the study, the results of this investigation showed a relatively high level of AST and ALT activities which is a consequent of tissue impairment due to heavy metal exposures.

Alkalinephosphatase (ALP) activity varied significantly between the two species with the activity higher in *C. nigrodigitatus* and lower in *C. senegalensis*. This may imply that *C. senegalensis* is less sensitive to heavy metals exposure compared to *C. nigrodigitatus*, since reduction in ALP activity may be taken as an index of hepatic parenchymal damage and hepatocytic necrosis (Onikienko, 1963; Kori-Siakpere et al., 2010). Reduced ALP activity by stressors probably indicates an altered transport of phosphate and an inhibitory effect on the cell growth and proliferation. Conversely, elevated plasma ALP activity as observed in *C. nigrodigitatus* may be as a consequence of osteoblastic activity increase or due to intra and extra hepatic obstruction of biliary passage (Jyotti and Narayanan 1999).

Blood parameters in fish have been studied to elucidate physiological adaptation and to assess the health of fishes (Vazquez and Guerrero 2007; Yildiz, 2009). Blaxhall and Diasley (1973) reported the use of haematocrit (PCV) and haemoglobin in checking anaemic conditions and reported values for fish usually vary from 20% to 35% (Pietse et al., 1981) and do not exceed 50% (Clarke et al., 1979).

The mean PCV value of 34.00% for *C. senegalensis* and 25.52% for *C. nigrodigitatus* recorded in the present study is within this range but this value for *C. nigrodigitatus* is lower than *C. nigrodigitatus* (35.52%) and *C. furcatus* (35.5%) from kwa River (Etim et al., 1999). The difference in mean haemoglobin, PCV and RBC values between *C. senegalensis* and *C. nigrodigitatus* could be attributed to differential responses caused by heavy metal exposures as it is well known that some metals cause early mortality of mature red blood cells and inhibition of haemoglobin formation through inhibition of erythrocyte alpha-amino levulinic acid dehydrogenase (ALA-D). Red blood cell (RBC) counts of  $2.71 \times 10^6 \text{ mm}^3$  for *C. senegalensis* and  $2.95 \times 10^6 \text{ mm}^3$  for *C. nigrodigitatus* was significantly high, according to Lenfant and Johnson (1972) erythrocyte counts greater than  $1 \times 10^6 \text{ mm}^3$  is considered high and indicative of high oxygen carrying capacity of the blood which is characteristic of fishes capable of aerial respiration and high metabolic rate. A strong inverse correlation ( $r=-0.94$ ) which occurred between haemoglobin and Iron in *C. nigrodigitatus* compared to a weak positive correlation ( $r=0.28$ ) recorded between haemoglobin and Iron in *C. senegalensis* could



indicate that haemoglobin increased with a fall in Iron and vice-versa in *C. nigrodigitatus* while there was a very slight increase in Iron with a corresponding slight increase in haemoglobin concentration, this observed relationship could imply that *C. nigrodigitatus* is more sensitive to Iron contamination compared to *C. senegalensis*. This is reflected in the slight difference between mean RBC count of *C. nigrodigitatus* ( $2.95 \times 10^6 \text{ mm}^3$ ) and *C. senegalensis* ( $2.7 \times 10^6 \text{ mm}^3$ ). The mean haemoglobin value of *C. senegalensis* ( $11.33 \pm 8.04 \text{ g/dl}$ ) and *C. nigrodigitatus* ( $13.32 \pm 8.29 \text{ g/dl}$ ) was high and fell within the range ( $5.6\text{--}15.8 \text{ g/dl}$ ) reported for *C. gariepinus* ( $8.70 \text{ g/dl}$ ) (Sowunmi, 2003). These values are also higher than  $4 \text{ g/dl}$  recorded for *Heterotis niloticus* (Fagbenro et al., 2000). This observed haemoglobin level may be connected to an adaptive response to heavy metal contamination or low dissolved oxygen recorded. The observed lymphocytes that were numerous than any other differential cells for *C. nigrodigitatus* ( $64.33 \pm 31.94\%$ ) and *C. senegalensis* ( $80.00 \pm 71.17\%$ ) and basophils that were seldomly seen are typical of most fishes (Ezzat et al., 1974; Kori-Siakpere and Ake 2005). Abundance of neutrophils in both species is similar to the observations of Breazile et al., (1982) and Kori-Siakpere and Ake (2005) and thought to be normal picture for fish blood. The major findings of this study are that heavy metals were accumulated by both species at concentration below WHO/FAO/FEPA permissible limits for human consumption. Heavy metals are toxic substances, with changes in enzymes activities in serum of fish exposed to various concentrations. Results of the study indicated that sub-lethal and chronic concentrations observed may cause changes in the haematological parameters and enzymatic activities of the studied fishes, hence could be used as biomarkers of metal contamination and detection in aquatic systems such as the Lagos Lagoon.

## CONCLUSION

Although results of heavy metal concentrations studied showed that consumption of the two species would not have any potential health implications; Similar studies may be performed to check contamination with other toxic metals such as mercury, cobalt, nickel and arsenic in commercial fish. Periodical monitoring of contaminants levels in fish by using enzymatic activities and haematological parameters in both commercial and feral fish is needful to checkmate any possible future accumulation and/ or increase as a result of waste and effluents discharge into the Lagoon. The use of feral fish species in local water bodies should be utilized as sentinel organisms for monitoring and management of the contamination status of these aquatic systems and should be

complementary with conventional physico-chemical water monitoring regimes.

## REFERENCES

- Abdelmeguid, N. Kheirallah, A.M. Abou-Shabana, K. and Abdel-Moneim, A. (2002). Histochemical and biochemical changes in liver of *Tilapia zill.G* as a consequence of water pollution. *J. Biol. Sci.* 2 (4): 224-229.
- Ademoroti, C.M.A. (1996). *Environmental Chemistry and Toxicology*, Foludex Press Ltd., Ibadan. 171 – 204.
- Adeyeye, E.I. Akinyugha, R.J. Febosi, M.E. and Tenabe, V.O. (1996). Determination of some metals in *Clarias gariepinus* (Cuvier and Valenciennes), *Cyprinus carpio* (L) and *Oreochromis niloticus* (L) fishes in a polyculture fresh water pond and their environment. *Aquacut.* 47. 205-214.
- Aliakbar, H. Alireza, S. Ahmad, S and Jasem, G.M. (2010). Assessment of Amino transferase enzymes in Yellow Sea Bream (*Acanthopagrus latus*) under experimental condition as Biomarkers of mercury pollution. *World. J.Fish.Mar.Sci.* 2(3): 196-192.
- Allen, P. (1995). Chronic Accumulation of Cadmium in the edible Tissues of *Oreochromis aureus* (Steindachner): Modification by Mercury and Lead. *Arch. Environ. Contam. Toxicol.* 29. 8-14.
- Babatunde, A.M. Waidi, O.A. and Adeolu, A.A. (2012). Bioaccumulation of heavy metals in fish: *Hydrocynus forskalii*, *Hyporhamphus bebe occidentalis* and *Clarias gariepinus* organs in downstream Ogun coastal water, Nigeria. *Transintl. J. Sci. Tech.* 2 (6): 119- 133.
- Blaxhall, P.C. and Daisley, K.W. (1973). Routine haematological methods for use with fish blood. *J. fish Biol.* 5: 771-781.
- Breazile, J.E. Zinn, L.L. Yauk, J.C, Mass, H.J. Wollscheid, J. (1982). A study of haematological profiles of channel catfish, *Ictalurus punctatus* (Rafinesque). *J. Fish. Biol.* 21:305–309.
- Clarke S, Whitmore, D.H. McMahon, R.F. (1979). Considerations of blood parameters of largemouth bass, *Micropterus salmoides*. *J. Fish. Biol.* 14:147–158.
- Don-Pedro KN, Oyewo OE, Otitoloju A.A. (2004). Trend measurement of heavy metals in Lagos lagoon ecosystems, Nigeria. *W. Afr. J. Appl. Ecol.*, 5: 103-114.
- Etim, L. Lebo, P.E. and King, R.P (1999). The dynamics of an exploited population of a silurid catfish (*Schilbe intermedius* Rüppell, 1832) in the Cross River, Nigeria. *Fisheries Res.*, 40: 295-307



- Ezzat, A.A, Shabana, M.B. Farghaly, A.M. (1974) Studies on the blood characteristics of *Tilapia zillii* (Gervias) I. Blood cells. *J. Fish. Biol.* 6:1-12.
- Fagbenro, O, Adedire, C.O. Ayotunde, E.O. Faminu, E.O. (2000). Haematological profile, food composition and enzyme assay in the gut of the African bony-tongue fish, *Heterotis (Clupisudis) niloticus* (Cuvier 1829) (Osteoglossidae). *Trop. Zool.* 13:1-9.
- Jyothi, B. and Narayan, G. (1999). Certain pesticide-induced carbohydrate metabolic disorders in the serum of freshwater fish *Clarias batrachus* (Linn.). *Food Chem. Toxicol.* 37: 417-421.
- Kalay, M. Aly. O. and Canil, M. (1999) Heavy metal concentrations in fish tissues from the Northeast Mediterranean sea. *Bull. Environ. Contam. Toxicol.* 63: 673-681.
- Karbassi, R. Bayati, I. Moattar, F. (2006). Origin and chemical partitioning of heavy metals in riverbed sediments. *Int. J. Environ. Sci. Tech.* 3(1): 35 – 42.
- Karthikeyan, S. Palaniappan, P.L.R.M. and Sabhanayakan, S. (2007) Influence of pH and water hardness upon Nickel accumulation in edible fish *Cirrhinus mrigala*. *J. Environ. Biol.* 28,484-492.
- Kori-Siakpere, O. and Ake, J.E.G. (2005) Blood cell types, morphology and dimensions of the African snakehead; *Parachanna obscura* (Osteichthyes: Channidae). *The Zoologist* 3:89-98.
- Kori-Siakpere, O. Ikom, R.B. and Ogbe, M. G. (2010). Variation in alanine aminotransferase activities in African Catfish: *Clarias gariepinus* at different sublethal concentrations of potassium permanganate. *Sci. Res. Essay* 5(12). 1501-1505.
- Kucuksezgin, F.A., Kontas, O., Altay, E. and Uluturhan, D. E. (2006). Assessment of marine pollution in Izmir Bay; Nutrient heavy metal and total hydrocarbon concentrations. *Environ. Int.* 32: 41-51.
- Lenfant, C. Johansen, K. (1972). Gas exchange in gill, skin and lung breathing. *Respir Physiol.* 14:211-218.
- Obasohan, E. E. (2008): The use of heavy metals load as an indicator of the suitability of the water and fish of ibiekuma stream for domestic and consumption purposes *Afri. J. of biotech.* vol. 7 (23), pp. 4345-4348
- Okoye, B.C.O. (1989). A Study of some heavy metals in Lagos Lagoon. *Ph.D Thesis*. Obafemi Awolowo University, Ile-Ife. 142pp.
- Olatunji, A.S. and Abimbola, A.F. (2010). Geochemical evaluation of the Lagos Lagoon sediments and water. *World. Appl. Sci. J.* 9 (2): 118-198.
- Onikienko, F. A., (1963) .Enzymatic changes from early stages of intoxication with small doses of chloroorganic insecticides. *Giginea. I. Fiziol. Ruda. Pro. Toksilol. Klinika* (Kietcv: Gos IZ. Med. Lit. 77p.
- Pietse, J.J. Smit, G.L. van Vleet, K.J, Schoobee, H.J, Hattingh, J. (1981). Some blood parameters of the Chinese grass carp, *Ctenopharygodon idella* (Valenciennes). *S. Afr. J. Zool.* 16(2):124-126.
- Roy, S.S (2002.). Some toxicological aspects of chlorpyrifos to the intertidal fish *Boleophthalmus dussumieri*. *Ph.D. Thesis*, University of Mumbai, India. 52-71.
- Singh, J, Hewawassam, H. Moffat, D. (1995). Nigeria: Strategic options for redressing industrial pollution. Industry and Energy Division, West Central Africa Department, 1: 45pp.
- Sowunmi, A.A. (2003). Haematology of the African catfish, *Clarias gariepinus* (Burchell, 1822) from Eleiyale reservoir, Ibadan, Southwest Nigeria. *The Zoologist* 2(1):40-44.
- Vazquez, G.R. Guerrero, G.A. (2007). Characterization of blood cells and haematological parameters in *Cichlasoma dimerus* (Teleostei, Perciformes). *Tissue and Cell* 39:151-160.
- Yang, H. and Rose, N.L. (2003). Distribution of Hg in the lake sediments across the UK. *Sci. Total Environ.* 304: 391 – 404.
- Yildiz, H.Y. (2009). Reference biochemical values for three cultured Sparid fish: striped sea bream, *Lithognathus mormyrus*; common dentex, *Dentex dentex*; and gilthead sea bream, *Sparus aurata*. *Comp Clin Pathol* 18:23-27.
- Younis, E.M. Abdel-Warith, A.A and Al-Asgah, N.A. (2012). Hematological and enzymatic response of Nile tilapia: *Oreochromis niloticus* during short and long term sublethal exposure to zinc. *Afr. J. Biotechnol.* 11 (19). 4442-4446.