# PHARMACOLOGICAL AND TOXICOLOGICAL ACTIVITIES OF THE METHANOLIC ROOT EXTRACT OF *CNESTIS FERRUGINEA* VAHL EX DE CANDOLLE (CONNARACEAE)

A THESIS SUBMITTED TO THE UNIVERSITY OF LAGOS, LAGOS, NIGERIA IN FULFILMENT OF THE REQUIREMENTS FOR THE AWARD OF DOCTOR OF PHILOSOPHY IN PHARMACOLOGY

By

ISHOLA, ISMAILA OGUNBAYODE

980702011

MAY, 2013

#### **DECLARATION**

This work titled ''Pharmacological and Toxicological activities of the methanolic root extract of *Cnestis ferruginea* Vahl ex De Candolle (Connaraceae)'' submitted to the School of Postgraduate Studies, University of Lagos, Lagos, Nigeria for the award of Doctor of Philosophy in Pharmacology was original research carried out by Ishola, Ismaila Ogunbayode in the Department of Pharmacology, College of Medicine of the University of Lagos, under the supervision of Professor (Mrs.) O.O. Adeyemi, Dr. (Mrs) E.O. Agbaje and Dr. Rakesh Shukla.

The work has not been submitted previously, in whole or in part, to qualify for any other academic award.

## ISHOLA, Ismaila Ogunbayode (Student)

## Professor (Mrs.) O.O. ADEYEMI (Supervisor)

Department of Pharmacology, Faculty of Basic Medical Sciences, College of Medicine of the University of Lagos, Idi-Araba, P.M.B. 12003 Lagos, Lagos, Nigeria.

## Dr (Mrs) E.O. AGBAJE (Supervisor)

Department of Pharmacology, Faculty of Basic Medical Sciences, College of Medicine of the University of Lagos, Idi-Araba, P.M.B. 12003 Lagos, Lagos, Nigeria.

### Dr SHUKLA, Rakesh (Supervisor)

Scientist F, Division of Pharmacology, Central drug Research Institute, (CSIR) Lucknow, India.

# **DEDICATION**

This thesis is dedicated to Almighty ALLAH- the most gracious and most merciful and, Mothers - I attribute all my success in life to the moral, intellectual and physical education I received from them.

#### **ACKNOWLEDGEMENTS**

My sincere appreciation goes to Professor (Mrs) Olufunmilayo Olaide Adeyemi for her immeasurable support, morally, materially, spiritually, financially, thorough supervision and guidance through the project. Words are not enough to quantify what God has used her to do in my life. Through her, I have been imbued with good scientific reasoning, organization and expression of research thoughts, and immense confidence in teaching Pharmacology.

I cherish the motherly guidance and support of my co-supervisor, Dr (Mrs) Esther Oluwatoyin Agbaje for her invaluable contributions. She is always willing to give audience and help proffer solutions to research challenges. She showed her care and concern at every point in time during this programme. My appreciation goes to my teachers at both the undergraduate and postgraduate levels Prof. S.A. Omilabu (Former Dean, Faculty of Basic Medical Sciences), Prof. A. Akintonwa, Dr. S.O. Olayemi, Dr. A.A. Akinyede and Dr. I.A. Oreagba who all have shaped my academic carrier at some point and they remain shining examples of intelligence, diligence and hard work.

I wish to appreciate the role Dr. A.J. Akindele and Dr. O. Awodele played in my life and in the course of this project from conception to birth; especially their contribution on data analysis, advice, wisdom, commitment and encouragement. I appreciate the support from Miss F.R. Aigbe, Mr. O.A. Salako, Mr. G.O. Afolayan, Dr. O. Nwaiwu, Dr. J.A. Kajero, Mr. S.O. Usman, Mrs. S.O. Amao (nee Lawal), and Mrs. O. Oyelakin of the Department of Pharmacology CMUL for their encouragement and being there for me when the occasion demanded.

My profound gratitude goes to Mr. Micah Chijioke (Department of Pharmacology), Mr. D.A. Ota, Mr. S. Dike (Department of Physiology), Mr. S.O. Adenekan (Department of Biochemistry), and Dr. D.I. Awelimobor (Department of Morbid Anatomy), all of the Faculty of Basic Medical Sciences, College of Medicine, University of Lagos, for the technical assistance given in the course of this research work. I also appreciate the effort of Mr. Anthony A. Ani of the AIDS Prevention Initiative Nigeria (APIN) Laboratory, Lagos University Teaching Hospital (LUTH), Lagos, Nigeria.

I wish to also express my appreciation to the technical staff of the Department of Pharmacology, including Mrs. V.C. Apugo, Mr. A.Y. Fashina, Mr. N. Nwose, Mr. O. Owagbayegun, and Mrs. R. Hussein for always being there to give their technical assistance

in the course of this programme. The administrative staff of the Department of Pharmacology from the 'HOA' Mrs. A.M. Akinremi, Mrs. C.N. Ashinze (my neighbour), Mrs. J.A. Oladubu (Retired), Mrs. T. Oderinde, and Mrs. S. Onetufo are appreciated for their help with administrative issues, goodwill and encouragement at all times. I also appreciate the support of my senior colleagues but very good friends with whom we rubbed minds, Dr. A.K. Oloyo (''ota mi''), Dr. W. Okunowo, Dr. A.O. Morakinyo, Dr. F.O. Awobajo, Dr. A.P. Arikawe, Mr. R. Lawal and Mr. S.I. Ogungbemi. I am immensely grateful to Mr. Joseph Ariwaodo of the Forestry Research Institute of Nigeria (FRIN), Ibadan, Oyo State, Nigeria, with whom I collected *Cnestis ferruginea*; and Professsor J.D. Olowokudejo of the Botany and Microbiology Department, University of Lagos, Nigeria, who identified and authenticated the plant. I got the first set of publications on *Cnestis ferruginea* from his shelves.

I am grateful to The World Academy of Sciences (TWAS) and Council of Scientific and Industrial Research, India (CSIR) for the TWAS-CSIR postgraduate fellowship which enabled me to conduct a 12 months research in the Division of Pharmacology, Central Drug Research Institute (CDRI), Lucknow, Uttar Pradesh, India. I acknowledge, Dr. Rakesh Shukla (my host supervisor at CDRI), for the scientific guidance, encouragement and above all his friendly attitude. It was this driving force that helped to achieve so much within the little time I spent. Dr T. Narender, Division of Medicinal Process Chemistry, guided me during the chromatographic and phytochemical elucidation processes. My sincere appreciation also goes to, Dr. (Mrs). Madhu Dikshit, (Head Division of Pharmacology), Dr. G. Palit (Scientist G), Dr. R. Ragubhir (Scientist G), Dr. C. Nath (Scientist G), Dr. Hanif Kashif, and Dr. (Mrs.) S. Shukla for their support during my stay at CDRI.

Dr. Surender Singh, Mr. Bharti Bhushan, Mr. C.P. Pandey, Mr. H.C. Verma, Mrs. Sachi Bharti, Mrs. S. Joshi and Mr. G.P. Singh are highly appreciated for their technical assistance. The technical support and moral support of Mr. J.P. Chatturvedi and Mrs. Rastogi Preeti during fractionation is highly appreciated.

I appreciate the technical support of Mrs. Manju Bhatnagar and Mrs. T.L. Seith during evaluation of cardiovascular and spasmogenic effects of the extract. God will see both of you through in CDRI.

I am highly grateful for the support and cooperation received from my colleagues in the Division of Pharmacology: (Santoshkumar Tota (Santy), Pradeep Kamat (PK), Shivika Rai, Subash Dwivedi, Niranjan Rajasekar, Ruby, Puneet, Vaibhav Tyagi, Madhu Kaundal,

Manavi Chatterjee, Neetu Singh, Pratibha Singh, Vaibhav Misra, Seema Singh, Vikas Kumar, Abhishek Desai, and Ankita Misra and Anubha Krishna) and Chemistry: (N. Rajender, G. Naresh (Nourish), Sriniwas Tiwari, Vinay Kumar Singh (VKS), Venkat, Satinath Sarkar, Madhu Gaurav, Shukla ('Bamakhan'), Faheem Khan, Preeti Rawat, Mameet Kumar, Preeti Dikshit, Saurav Bera, Sarkanlan, Ritish, Imran, Kamil, Jamal, Dinesh Yadav and many others that made my stay in Lucknow a memorable one.

I appreciate the moral support of all the TWAS fellows in Lucknow; Dr. Hassanwara Sanusi, Dr. G. Adekunle, Dr. Charles Otieno, Mrs. Olajumoke Ojo (May the Good Lord strengthen you and give you the fortitude to bear the irreparable loss of your husband), Miss. Arlette Vyry, Miss. Joy Odimegwu and Mr. A. Boniface.

The sincere concern, spiritual and financial support of my mother from cradle to adulthood since the demise of my father can never be forgotten, YOU ARE INDEED A MOTHER (Mrs. Maryam Adeniyi), May ALLAH (SWT) preserve and protect you to reap the fruit of your hard labour and may He grant you all the goodness of this world and hereafter and tranquillity (Amen). In the course of this programme I lost my only brother Idris who is a sickle cell patient may ALLAH (SWT) grant him Al Jannah Firdaus.

To my siblings Rasheedah Sholaja, Bukola Adeniyi and Olabisi Adeniyi, I say a big thank you for your moral and spiritual support. My very big Sister (Mrs. Olufunmilayo Adepoju) who stood by my family while I was away from the country, thank you for being there for my family and never failing to help out when the occasion demanded.

To my dearest friend and better half, Mrs. Halimah Ishola for her remarkable understanding, spiritual, moral, emotional support, and perseverance when most needed and absorbing all the pressure of the critical periods of this work. I acknowledge the support of master Abdulahi **Ishola** (my son) who atimes stayed with me in the office till 12 midnight, many at times both of us being locked up in the Department and to my lovely jewel Nafisah Motunrayo Ishola for her support taking care of her mum while I was away. I appreciate all that contributed to this work, not mentioned.

Finally, all glory, exaltation and adoration belong to Almighty Allah, the Lord of the whole universe for his protection, mercies, wisdom, and strength, without whom there would not be me and this thesis.

# TABLE OF CONTENTS

DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	vii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
HISTOLOGIC SLIDES	xviii
ABSTRACT	xix
1.0 INTRODUCTION	2
1.1 Background of study	2
1.2 Statement of problem	4
1.3 Objectives of study	6
1.4 Significance of study	6
1.5 Operational definition of terms	7
1.6 List of abbreviations	9
2.0 LITERATURE REVIEW	11
2.1 Overview of inflammation	12
2.2 Evidence for the contribution of inflammation to CNS disease	14
2.2.1 Epilepsy	15
2.2.2 Alzheimer's disease	15
2.3 Pain	16
2.4 Therapeutic management of inflammation and pain	18
2.4.1 Therapeutic uses of NSAIDs	20
2.4.1.1 Adverse effects of NSAIDs	21

	2.4.2 Opioid Analgesics	21
	2.4.2.1 Opioid receptors	22
	2.4.2.2 Adverse effects of opioid analgesics	23
	2.4.3 Glucocorticoids: anti-inflammatory and immunosuppressive agents	23
	2.4.3.1 Toxicity of adrenocortical steroids	24
	2.4.3.2 Withdrawal of therapy	24
	2.4.3.3 Continued use of supraphysiological glucocorticoids doses	25
2.5 De	pression and Anxiety	25
	2.5.1 Depression	26
	2.5.1.1 Experimental models used in the evaluation of potential	27
	antidepressants.	
	2.5.1.2 Antidepressants	28
	2.5.2 Anxiety	29
	2.5.2.1 Pharmacotherapy of anxiety	30
2.6 Ov	erview of epilepsy	32
	2.6.1 Pathophysiology of epilepsy	32
	2.6.2 Nature and mechanisms of seizures	33
	2.6.3 Pharmacological treatment of epilepsy	34
2.7 Pat	chophysiology of Alzheimer's disease	35
	2.7.1 Rationale for disease modifying strategies	35
	2.7.2 Oxidative stress in Alzheimer's disease	37
	2.7.1.1 Antioxidants for prevention of Alzheimer's disease	38
	2.7.2 Treatment of Alzheimer's disease	38
2.8 Pla	ant description	39
	2.8.1 Botanical profile	39
	2.8.2 Range	40
	2.8.3 Local names in Nigeria	40
	2.8.4 Phytochemistry	41
2.9 Eth	nnobotanical uses	41
2.10 Pl	harmacological studies	41
	2.10.1 Antimicrobial activity	41
	2.10.2 Antioxidant activity	42
	2.10.3 Hypoglyceamic activity	42

3.0 MATERIALS AND METHODS	43
3.1 Plant materials	44
3.2 Preparation of plant extract	44
3.3 Bioactivity guided fractionation	44
3.3.1 General experimental procedure	45
3.4 Laboratory animals	45
3.5 Toxicological evaluation	46
3.5.1 Acute toxicity test	46
3.5.2 Sub-chronic toxicity test	46
3.5.3 Effect on vital organs	47
3.5.4 Measurement of in vivo antioxidants and MDA levels	47
3.5.5 Histological assessment	47
3.5.6 Haematological assessment	47
3.5.7 Biochemical assessment	48
3.5.8 Sperm analysis	48
3.5.9 Sperm motility	48
3.5.10 Sperm count	48
3.5.11 Sperm morphology	49
3.6 Pharmacological procedure	49
3.6.1 Analgesic activity	49
3.6.1.1 Mouse writhing test	49
3.6.1.2 Formalin test	49
3.6.1.3 Haffner's tail clip test	50
3.6.1.4 Hot plate test	50
3.6.1.5 Investigation of possible mechanism(s) of analgesic effect	51
3.6.2 Anti-inflammatory activity	52
3.6.2.1 Carrageenan-induced paw oedema test	52
3.6.2.2 Egg albumin-induced paw oedema test	52
3.6.2.3 Histamine- and serotonin-induced paw oedema test	52
3.6.2.4 Xylene-induced ear oedema test	53
3.6.2.5 Formaldehyde-induced arthritis inflammation test	53
3.6.3 Anticonvulsant activity	54
3.6.3.1 Maximal electroshock-induced seizure test	54
3.6.3.2 Strychnine-induced seizure test	54

3.6.3.3 Picrotoxin-induced seizure test	54
3.6.3.4 Bicuculline-induced seizure test	55
3.6.3.5 Isoniazid-induced seizure test	55
3.6.3.6 Yohimbine-induced seizure test	55
3.6.4 Behavioural observations	55
3.6.4.1 Spontaneous motor activity	56
3.6.4.2 Rotarod test	56
3.6.5 Antidepressant activity	57
3.6.5.1 Forced swimming test	57
3.6.5.2 Tail suspension test	57
3.6.5.3 Elucidation of mechanism(s) of antidepressant activity	57
3.6.6 Anxiolytic activity	58
3.6.6.1 Hole board test	58
3.6.6.2 Elevated plus maze test	59
3.6.6.3 Light-dark test	59
3.6.6.4 Elucidation of mechanism(s) of anxiolytic activity	59
3.6.7 Antidementic activity	
3.6.7.1 Drugs and treatment regimens	60
3.6.7.2 Administration of scopolamine	60
3.6.7.3 Passive avoidance test	60
3.6.7.4 Morris water maze test	61
3.6.7.5 Estimation of biochemical parameters	61
3.6.7.6 Brain tissue preparation	61
3.6.7.7 Acetylcholinesterase assay in the brain	62
3.6.7.8 Measurement of malondialdehyde	62
3.6.7.9 Measurement of glutathione	62
3.6.7.10 Measurement of nitrite	62
3.6.7.11 Protein estimation	63
3.7 <i>In vitro</i> study	63
3.7.1 Cell culture	63
3.7.2 MTT assay for cytotoxicity assessment	63
3.7.3 Estimation of nitrite level	64
3.7.4 Measurement of reactive oxygen species generation	64
3.7.5 Estimation of glutathione level	65

	3.7.6 Estimation of malondialdehyde level	65
	3.7.7 Estimation of TNF- $\alpha$ in THP-1 cells	66
	3.7.7.1 Stimulation of cells and collection of supernatants	66
3.8 S	tatistical analysis	66
3.9 D	Orugs, chemicals and reagents	67
4.0 R	ESULTS	68
4.1 T	oxicity studies of C. ferruginea	69
	4.1.1 Acute toxicity study	69
	4.1.2 Mortality in the subchronic toxicity test	72
	4.1.3 Body weight	72
	4.1.4 Sperm parameters	74
	4.1.5 Effect on vital organs	74
	4.1.6 In vivo antioxidant enzymes and MDA	79
	4.1.7 Haematological parameters	81
	4.1.8 Biochemical parameters	81
	4.1.9 Histological assessment	86
	4.1.9.1 Heart	86
	4.1.9.2 Kidney	86
	4.1.9.3 Liver	86
	4.1.9.4 Testes	88
	4.1.9.5 Ovary	88
	4.1.9.6 Reversibility study	90
4.2 A	analgesic activity	93
	4.2.1 Acetic acid-induced writhing test	93
	4.2.2 Formalin test	96
	4.2.3 Haffner's tail clip test	98
	4.2.4 Hot plate test	100
4.3 A	anti-inflammatory activity	103
	4.3.1 Carrageenan-induced paw oedema test	103
	4.3.2 Egg albumin-induced paw oedema test	108
	4.3.3 Histamine-induced paw oedema test	108
	4.3.4 Serotonin-induced paw oedema test	108
	4.3.5 Xylene-induced ear oedema test	112
	4.3.6 Formaldehyde-induced arthritis inflammation test	114

4.4 Anticonvulsant activity	116
4.4.1 Maximal electroshock-induced seizure	116
4.4.2 Strychnine-induced seizure	116
4.4.3 Picrotoxin-induced seizure	119
4.4.4 Bicuculline-induced seizure	119
4.4.5 Isoniazid-induced seizure	122
4.4.6 Yohimbine-induced seizure	122
4.5 Spontaneous motor activity	125
4.6 Rotarod test	125
4.7 Antidepressant activity	127
4.7.1 Forced swimming test	127
4.7.2 Tail suspension test	130
4.8 Anxiolytic activity	132
4.8.1 Hole board test	132
4.8.2 Elevated plus maze test	132
4.8.3 Light-dark test	132
4.8.3 Light-dark test 4.9 Antidementic activity 4.9.1 Passive avoidance test	
4.9.1 Passive avoidance test	136
4.9.2 Morris water maze test	138
4.9.3 Acetylcholinesterase activity	142
4.9.4 Oxidative stress parameters	144
4.9.4.1 Malondialdehyde level	144
4.9.4.2 Glutathione level	144
4.9.4.3 Nitrite level	144
4.9.5 <i>In vitro</i> anti-inflammatory activity of CF-2 and CF-5 in C6 cell line	147
4.9.5.1 MTT cell viability assay	147
4.9.5.2 LPS-induced nitrite release in C6 cells	147
4.9.5.3 LPS-induced ROS generation in C6 cells	147
4.9.5.4 LPS-induced MDA formation in C6 cells	147
4.9.5.5 LPS-induced GSH release in C6 cells	152
$4.9.5.6$ LPS-induced TNF- $\alpha$ release in THP-1 cells	152
5.0 DISCUSSION	156
5.1 Toxicological evaluation	156
5.1.2 Acute toxicity test	156

5.1.3 Subchronic toxicity test	156
5.2 Analgesic activity	160
5.3 Anti-inflammatory activity	162
5.4 Bioactivity guided isolation of analgesic and anti-inflammatory constituents	163
5.5 Anticonvulsant activity	164
5.6 Antidepressant activity	165
5.7 Anxiolytic activity	168
5.8 Antidementic activity	170
5.9 Effect on pro-inflammatory markers	172
5.10 Phytochemical constituents	173
6.0 CONCLUSIONS	173
7.0 SUMMARY OF FINDINGS	174
7.0 CONTRIBUTIONS TO KNOWLEDGE	176
8.0 REFERENCES	177
9.0 APPENDIX	212

# LIST OF TABLES

Table 1: Effect of <i>C. ferruginea</i> on body weight of rats	73
Table 2A: Effect of <i>C. ferruginea</i> on sperm parameters and change in body weight	75
Table 2B: Effect of <i>C. ferruginea</i> on sperm parameters (reversibility)	76
Table 3A: Effect of <i>C. ferruginea</i> on organ weight	77
Table 3B: Effect of <i>C. ferruginea</i> on organ weight (reversibility study)	78
Table 4: Effect of <i>C. ferruginea</i> on <i>in vivo</i> antioxidant enzymes and MDA level	80
Table 5A: Effect of <i>C. ferruginea</i> on haematological parameters	82
Table 5B: Effect of <i>C. ferruginea</i> on haematological parameters (reversibility)	83
Table 6A: Effect of <i>C. ferruginea</i> on serum biochemical parameters	84
Table 6B: Effect of <i>C. ferruginea</i> on serum biochemical parameters (reversibility)	85
Table 7: Elucidation of the mechanism of analgesic effect in mouse writhing test	95
Table 8A: Effect of <i>C. ferruginea</i> on formalin-induced pain	97
Table 8B: Further elucidation of the mechanism of analgesic effect in formalin test	97
Table 9: Effect of <i>C. ferruginea</i> on tail clip-induced pain	99
Table 10A: Effect of <i>C. ferruginea</i> on hot plate test	101
Table 10B: Effect of amentoflavone on hot plate test	102
Table 11A: Effect of <i>C. ferruginea</i> on carrageenan-induced paw oedema	104
Table 11B: Effect of <i>C. ferruginea</i> fractions against carrageenan-induced paw oedema	105
Table 11C: Effect of <i>C. ferruginea</i> subfractions against carrageenan-induced paw edema	106
Table 11D: Effect of CF-2 and CF-5 against carrageenan-induced paw oedema	107

Table 12: Effect of <i>C. ferruginea</i> on egg albumin-induced paw oedema	109
Table 13: Effect of <i>C. ferruginea</i> on histamine-induced paw oedema	110
Table 14: Effect of <i>C. ferruginea</i> on serotonin-induced paw oedema	111
Table 15: Effect of <i>C. ferruginea</i> on xylene-induced ear oedema	113
Table 16: Effect of <i>C. ferruginea</i> on formaldehyde-induced arthritis inflammation	115
Table 17: Effect of <i>C. ferruginea</i> on maximal electroconvulsive shock	117
Table 18: Effect of <i>C. ferruginea</i> on strychnine-induced seizure	118
Table 19: Effect of <i>C. ferruginea</i> on picrotoxin-induced seizure	120
Table 20: Effect of <i>C. ferruginea</i> on bicuculline-induced seizure	121
Table 21: Effect of <i>C. ferruginea</i> on isoniazid-induced seizure	123
Table 22: Effect of <i>C. ferruginea</i> on yohimbine-induced seizure	124
Table 23: Elucidation of mechanism of anxiolytic effect in EPM	135

# LIST OF FIGURES

Figure 1: Schematic representation of fractionation procedure	44
Figure 2: Median lethal dose (LD <sub>50</sub> ) following oral administration	70
Figure 3: Median lethal dose (LD <sub>50</sub> ) following intraperitoneal administration	71
Figure 4: Effect of CF extract, fractions, CF-2 and CF-5 on mouse writhing	95
Figure 5: Time course effect of CF on spontaneous locomotor activity	126
Figure 6A: Effect of CF on forced swimming test	128
Figure 6B: Effect of CF-2 on forced swimming test	128
Figure 6C-E: Elucidation of mechanism(s) of CF antidepressant activity	128
Figure 7 A-F: Elucidation of mechanism(s) of CF-2 antidepressant activity	129
Figure 8 A-D: Effect of CF and CF-2 on tail suspension test	131
Figure 9A: Effect of CF and CF-2 on hole board test	134
Figure 9B: Effect of CF and CF-2 on elevated plus maze test	134
Figure 9C: Effect of CF and CF-2 on light-dark compartment test	134
Figure 10A: Effect of CF on passive avoidance test	137
Figure 10B: Effect of CF-2 and CF-5 on passive avoidance test	137
Figure 11A-D: Effect of CF on Morris water maze test	139
Figure 12A-C: Effect of CF-2 and CF-5 on Morris water maze test	139
Figure 13A: Effect of CF on acetylcholinesterase activity	143
Figure 13B: Effect of CF-2 and CF-5 on acetylcholinesterase activity	143
Figure 14A: Effect of CF on malondialdehyde level	145
Figure 14B: Effect of CF-2 and CF-5 on malondialdehyde level	145
Figure 14C: Effect of CF on glutathione level	145

Figure 14D: Effect of CF-2 and CF-5 on glutathione level	145
Figure 15A: Effect of CF on nitrite level	146
Figure 15B: Effect of CF-2 and CF-5 on nitrite level	146
Figure 16A-B: Effect of CF-2 and CF-5 on cell viability in C6 cell line	148
Figure 17A-B: Effect of CF-2 and CF-5 on nitrite release in C6 cell line	149
Figure 18A-B: Effect of CF-2 and CF-5 on reactive oxygen species generation	150
Figure 19A-B: Effect of CF-2 and CF-5 on malondialdehyde formation in C6 cell line	151
Figure 20A-B: Effect of CF-2 and CF-5 on glutathione deficit in C6 cell line	153
Figure 21: Effect of CF-2 on TNF-α release in THP-1 cell line	154

# HISTOLOGIC SLIDES

Plate 1: Histologic presentation of rat liver in control group	87
Plate 2: Histologic presentation of rat liver in CF 80 mg/kg group	87
Plate 3: Histologic presentation of rat liver in CF 400 mg/kg group	87
Plate 4: Histologic presentation of rat liver in CF 1000 mg/kg group	87
Plate 5: Histologic presentation of rat testes of control group	89
Plate 6: Histologic presentation of rat testes in CF 1000 mg/kg group	89
Plate 7: Histologic presentation of rat kidney in CF 80 mg/kg group (reversibility)	91
Plate 8: Histologic presentation of rat liver in CF 1000 mg/kg group (reversibility)	92
Plate 9: Histologic presentation of rat testes in CF 80 mg/kg group (reversibility)	93

#### **ABSTRACT**

Cnestis ferruginea Vahl ex DC (Connaraceae) (CF) is a shrub widely used in traditional African medicine for the treatment of various painful inflammatory conditions and psychiatric disorders. The study was undertaken to investigate the toxicological profile and pharmacological effects with a view to isolate and characterize the active constituents of the methanolic root extract of CF responsible for these effects. Acute toxicity tests were carried out in mice and median lethal dose (LD<sub>50</sub>) was estimated following oral and intraperitoneal administrations and as basis for dose selection in the pharmacological studies. Subchronic toxicity (90 days (plus 14 days reversibility)) studies were conducted in rats with oral daily doses of 100, 400 and 1000 mg/kg. Parameters observed for at the end of chronic tests include changes in body weight and vital organs weight, mortality, heamatological, biochemical, histological and oxidative stress parameters. Analgesic activity of the extract (100-400 mg/kg, p.o.) was evaluated using the acetic acid-induced writhing, formalin, tail clip and hot plate tests. The possible mechanism of its analgesic effect was investigated in mouse writhing and formalin tests using naloxone ( $\mu$ - opioid antagonist), prazosin ( $\alpha_1$ adrenoceptor antagonist), ondansetron (5- HT<sub>3</sub> antagonist), haloperidol (D<sub>2</sub>- receptor antagonist) and glibenclamide (ATP sensitive potassium channels). The anti-inflammatory effect was investigated using carrageenan-, egg albumin-, serotonin-, histamine-, and formaldehyde-induced rat paw oedema and xylene-induced ear oedema. Separation of phytochemical constituents and bioactivity guided assays were carried out with fractions on mouse writhing, and hot plate tests (analgesic effect) and carrageenan-induced rat paw oedema (anti-inflammatory effect). The maximal electroconvulsive (MES)-, strychnine-, picrotoxin-, bicuculline-, isoniazid-, and yohimbine-induced seizures were used to evaluate anticonvulsant effect in mice. The spectrum of activities of the extract on psychiatric disorders was studied using forced swimming and tail suspension tests to evaluate antidepressant effect while hole board, elevated plus maze (EPM), and light-dark tests were used to investigate anxiolytic effect. The involvement of 5-HT<sub>2</sub> receptor,  $\alpha_1$ - and  $\alpha_2$ adrenoceptor, dopamine D<sub>2</sub> receptor, muscarinic cholinergic receptor, and nitric oxide pathway in the antidepressant effect of CF and CF-2 were investigated. Antidementic activity of the extract and its constituents were investigated using scopolamine-induced memory deficit in mice. Memory function was evaluated by passive avoidance and Morris water maze tests. Biochemical parameters of oxidative stress and cholinergic function were also estimated. The in vitro study was carried out to investigate the effect of CF-2 and CF-5 on

neuroinflammatory markers (oxidative stress, nitrative stress, and tumour necrotic factoralpha (TNF-α)) in C6 and THP-1 cells respectively. Acute oral and intraperitoneal administrations of CF (>2 g/kg and >400 mg/kg respectively) produced behavioural signs of toxicity as well as mortalities within 24 h with estimated LD<sub>50</sub> of 5.22 g/kg (p.o.) and 643.65 mg/kg (i.p.). In the subchronic test, CF at 100 mg/kg did not produce any significant irreversible deleterious effect on weight of animals and vital organs, in vivo antioxidants, heamatological, biochemical, sperm parameters and histological presentation. Platelet anomaly was elicited as delayed effect. The effects of the extract at 400 and 1000 mg/kg were similar but with delayed anaemia in females and weight reduction in males as side-effects. CF generally showed a potential to induce in vivo antioxidants enzymes. The methanolic root extract of Cnestis ferruginea (100, 200, and 400 mg/kg; p.o.) produced significant (P < 0.05) dose-dependent inhibition of pain response elicited by acetic acid and formalin while also increasing the nociceptive reaction latency in the tail clip and hot plate tests. The analgesic activity of the extract was significantly (P<0.01) reversed following naloxone, yohimbine, ondansetron, haloperidol, and glibenclamide pretreatment. In respect of anti-inflammatory activity, Cnestis ferruginea caused significant (P < 0.05) dose-dependent inhibition of oedema development in the carrageenan, egg albumin, serotonin, histamine, formaldehyde, and xylene-induced inflammation tests. The effects of the extract in the various models were generally comparable to those of the standard drugs used. Due to the promise shown by CF, an activity guided-isolation of active constituents was carried out which led to the isolation of amentoflavone (CF-2; a bioflavonoid) and an amino acid-like compound (CF-5) through column chromatography and spectroscopic methods (<sup>1</sup>H NMR, <sup>13</sup>C NMR and HMBC). The methanolic root extract of Cnestis ferruginea (50-400 mg/kg, p.o.) produced significant (P <0.05) antagonism of MES-induced seizures and ameliorated the seizure induced by strychnine with peak effect observed at 50 mg/kg. CF produced significant (P < 0.05) increase in onset of tonic convulsion which was comparable to the effect of clonazepam (0.5 mg/kg, p.o.). The extract produced 40, 20 and 20% protection respectively at 100, 200 and 400 mg/kg in picrotoxin-induced seizure. CF (100-400 mg/kg) completely antagonized bicuculline-induced seizure. Similarly, the extract produced dose-dependent increase in percentage protection in isoniazid and yohimbine-induced seizure in mice. Acute treatment with CF and its constituents significantly (P < 0.001) reduced the duration of immobility dose dependently in FST and TST. The pretreatment of mice with metergoline (4 mg/kg, i.p., a 5-HT2 receptor antagonist) and reserpine (2 mg/kg, i.p., a drug known to induce depletion of biogenic amines) 15 mins before the administration of CF (100 mg/kg; p.o.) significantly

prevented its antidepressant effect in the FST. However, pretreatment with prazosin (62.5  $\mu g/kg$ , i.p., an  $\alpha_1$ -adrenoceptor antagonist), yohimbine (1 mg/kg, i.p., an  $\alpha_2$ -adrenoceptor antagonist), sulpiride (50 mg/kg, i.p., a dopamine D<sub>2</sub> receptor antagonist), atropine (1 mg/kg, i.p., a muscarinic receptor antagonist) did not prevent this effect. The extract and its components produced significant (P < 0.05) attenuation of anxiety shown by the increased number of head-dips in hole board, increased time spent in the open arms in EPM and increased exploration of light chamber. The anxiolytic effect of CF and CF-2 were reversed by flumazenil (3 mg/kg, i.p.) pretreatment. Scopolamine (3 mg/kg, i.p.) produced a decrease in transfer latency time (TLT) and an increase in escape latency time (ELT) in passive avoidance and Morris water maze tests respectively which are signs of memory deficits along with increased acetylcholinesterase (AChE) activity and oxidative stress in mice brain. Oral administration of CF, CF-2 and CF-5 significantly (P < 0.05) reversed scopolamine-induced memory impairments shown by the increased transfer latency time in passive avoidance test and decreased escape latency time in Morris water maze test. They also significantly (P <0.05) inhibited AChE and enhanced antioxidant enzyme activities in the brain following scopolamine injection as compared to vehicle administration in scopolamine (i,p)-treated mice which was comparable to effect of tacrine. Lipopolysaccharide (LPS) (10 µg/ml) stimulated C6 cells to release nitrite, reactive oxygen species (ROS), and malondialdehyde (MDA) while it down regulated glutathione (GSH) in C6 cells. Similarly, LPS up regulated the release of TNF- $\alpha$  in THP-1 cells. However, CF-2 and CF-5 significantly (P<0.001) attenuated nitrite release, ROS generation, MDA level and also up regulated the level of GSH. In addition, produced significant (P<0.05) attenuation of TNF-  $\alpha$  level. CF-2 and CF-5, per se treatment did not have any significant effect on C6 and THP-1 cells. In conclusion, the methanolic root extract of *Cnestis ferruginea* given over an extended period is relatively safe and possesses significant analgesic, anti-inflammatory, anticonvulsant, antidepressant, anxiolytic, and antidementic effects, and these effects might have been produced by amentoflavone and/ amino acid like compound (CF-5) or combination of the phytoconstituents.