

## **Plankton and Microbenthos Communities of Freshwater Habitats in Kogi State, North-Central Nigeria.**

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### **Abstract**

Three different rivers the Okpo, Idebu and parts of the Niger River were investigated for both micro and macro biota. The pH of the water showed neutral (7.07) water body with salinity of 0.03% and high dissolved oxygen content (5.1 mg/l). In the dry season, sulphate, Phosphate-phosphorus and Nitrate-nitrogen recorded 3.0, 0.003 and 0.138 mg/l respectively while 2.0, 0.005 and 0.210 mg/l were recorded in the wet season. The divisions, Bacillariophyta, Chlorophyta and Cyanophyta were recorded with pennate diatoms dominating the phytoplankton spectrum. The phyla; Cladophora, Copepoda and Rotifera represented the zooplankton with rotifers being the frequently encountered group. The macrobenthos recorded were Mollusca, Insecta, Hirudinea, Crustacea and Oligochaeta. High species in terms of number were recorded in wet season; this may be a pointer to environmental conditions such as rainfall and nutrients as major factors controlling the plankton abundance. Community structure analysis showed a relatively stressed environment.

### **Introduction**

Plankton are the starting point of energy transfer in the aquatic ecosystem and are very sensitive to the environment they live in and any alteration in the environment leads to the change in the plankton communities in terms of

tolerance, abundance, diversity and dominance in the habitat. Phytoplankton represent the base of food webs in any ecosystem and play a major role in the global cycling of carbon, nitrogen, phosphorus and other elements and the regulation of earth's climate. The biomass distribution and species composition of phytoplankton have important effects on carbon fixation rates and on transfer of energy in food webs. Therefore, plankton population observation may be used as a reliable tool for biomonitoring studies to assess the pollution status of aquatic bodies (Mathivanan and Jayakumar, 1995). The knowledge of plankton species composition and distribution to time and space are of great value especially in any running water system. As growing populations, progressive industrialization and intensification of agriculture are leading to increased pollution of our surface waters, there is a need for conservation of our resources especially water. According to Biddanda and Benner (1997), one of the main sources of carbohydrates in many aquatic systems is phytoplankton, where biomass is typically made up of from 15 to 35% carbohydrate. Townsend *et al.* (2000) stated that the distribution, abundance, species diversity and composition of the phytoplankton can be used to assess the biological integrity of a water body. Peerapornpisal *et al.* (2004) opined that species composition of the phytoplankton community is an efficient bioindicator for water quality. In Nigeria, some recent phytoplankton and macrobenthic fauna work are those of Abdullahi and Indobawa (2005) who worked on the phytoplankton of Hadeija river, Jigawa; Abubakar (2007) reported phytoplankton community of Lake Nguru, Kano; Adesalu and Nwankwo (2005, 2008) reported on Olero and Abule-Eledu creeks respectively; Adesalu *et al.* (2008) worked on Ogbe creek; Aneni and Hassan (2003) observed the abundance of plankton at Kudeti and Onireke stream Ibadan; Chindah and Pudo (1991) worked on Bonny river, Niger Delta; Dimowo (2013) reported on River Ogun; Edokpayi *et al.*, investigated macrobenthic fauna of Kuramo water, Lagos; Ekwu and Sikoki (2005) worked on Cross River estuary; Emmanuel and Onyema (2007) investigated the Abule-Agege creek; New Calabar river phytoplankton was analysed by Erundu and Chindah (1991), Imoobe (2011) observed the Okhuo river while Mohammad and Saminu (2012) gave report on Salanta River Kano. Studies of the abundance, distribution and

composition of phytoplankton communities are, therefore, a fundamental contribution to our understanding of the structure and function of ecosystems hence this study was carried out to know the status of these water bodies using plankton and microbenthos as indicators of the environment.

## **Materials and Methods**

### **Description of study sites**

Kogi State is situated in the savannah region with the State capital at Lokoja, located at the confluence of the two major Rivers, Niger and Benue. The State is bordered to the East by Benue State, to the South by Edo, Enugu and Ondo States, to the North by Niger and Nasarawa States and to the West by Kwara and Ekiti States. There are two main seasons, dry and wet. The wet season begins in May and stops towards the end of October. The ranges in mean monthly minimum and maximum temperatures during wet and dry seasons in the region were 21.9-26.5 and 30.7-36.7°C, respectively.

**Okpo River** a seasonal river, has an average width of about 6m with an average depth of about 0.3m during the dry season. Its depth increases to a maximum of about 1.2m during the wet season. It drains into River Niger.

**Idebu River** also a seasonal river has an average width of about 6.3m with average depth of about 0.5m during the dry season. Its depth increases to a maximum of about 1.2m during the wet season. The two river systems have no significant flow during dry season. It drains into River Niger.

**River Niger:** The River Niger, according to Goulden and Few (2011) is a transboundary river, originating from Guinea, flowing through Mali, Niger, forming the border with Benin and discharging into the Atlantic Ocean in Nigeria. The River Niger is a source of drinking water, hydro-electric power, irrigation, transportation, fishing among other uses (Ali *et al.*, 2013). The study was carried out in the portion of the lower River Niger that falls

in Ofu local government area of the State and its geographical coordinate at the access point is given as  $07^{\circ} 24' 46.518''\text{N}$ ,  $06^{\circ} 44' 42.081''\text{E}$  which is approximately 60km to Lokoja, 260km to Abuja, approximately 120km to Kabba.



Part of Idebu River at Itobe

Section of River Niger at Itobe

### **Collection of samples**

**Plankton:** Samples were collected twice to denote the seasons (Wet and Dry). Due to the shallowness of the study site, a water sampler was used to collect the water samples directly which were transferred into well labeled 500ml plastic containers with screw caps and preserved with 4% unbuffered formalin. Surface water samples for physico-chemical analysis were collected in 1.5ml containers. Some analyses (pH and salinity) were done in-situ before taking the water samples in an ice chest to the laboratory for further analysis. Quantitatively, various plankton species were computed by calculating average cells ( $\pm 5\%$ ) per transect following the method described by Vollenweider (1996). All identifications of plankton taxa were done using keys and illustrations presented in Davis (1955); Desikachary (1959); Hendey (1964); Patrick and Reimer (1966, 1975); Olaniyan (1968, 1969, 1975); Wiafe and Frid (2001); Hustedt (1930–1937) Whitford and Schumacher (1973); Krammer and Large-Bertalot (1986); Adesalu 2008; Nwankwo (1984) and Wimpenny (1966).

**Macrobenthos:** At each station, soil samples were scooped directly at the sampling points due to shallowness of the sites. The sediments were sieved

through a 0.5mm mesh size stainless sieve. The sieve and its contents were immersed in the native water and gently agitated until organisms debris matrix were removed. The sieve contents were transferred into properly labeled, wide mouthed glass jars containing 4% unbuffered formalin solution for preservation. The macrobenthos was observed under the Olympus microscope at different magnifications and biological species documented. Identifications were made using appropriate keys (Needham and Needham, 1962; Quigley, 1977; Atobatele, *et al.*, 2005; Ibemenuga and Inyang, 2006). All bio-statistical methods used were according to Ogbeibu (2005).

**Physico-chemical analysis:** Surface water samples were analysed using APHA 1998 methods. Air and water temperatures were measured in-situ using mercury in glass thermometer while the pH was determined with Electronic Cole Parmer Testr3. Salinity was measured using a handheld refractometer. Gravimetric method was applied for both total dissolved and suspended solids. Dissolved oxygen and chemical oxygen demand were determined using titrimetric method while Winkler's method was applied for the determination of biological oxygen demand. Colorimetric method was applied for the nutrients contents (nitrate-nitrogen and phosphate-phosphorus) while turbidimetric method was used for sulphate (APHA 1998).

**Community structure analysis:** To obtain the estimate of species diversity, three community structure indices were used: Margalef's diversity index (d), Shannon-Weaner Index ( $H^1$ ) (Shannon & Weaver 1963) and Species Equitability (j) or Evenness (Pielou 1975).

## Results

### Physico-chemical

The pH value was 7.07 in the dry season and 7.54 was observed in wet month while the salinity was between 0.02%. - 0.03%. In the dry season, Sulphate, Phosphate-phosphorus and nitrate-nitrogen recorded 3.0, 0.003 and 0.138 mg/l respectively while 2.0, 0.005 and 0.210 mg/l were recorded

in the wet season. Highest value (24.0 mg/l) of total suspended solids was measured in dry season while 38.1 mg/l was observed for total dissolved solids in wet months. Oil and grease were detected only in wet season with 0.02 mg/l recorded. Dissolved oxygen value was high with 5.1mg/l and 4.5mg/l recorded in dry and wet seasons respectively while biological oxygen demand and chemical oxygen demand recorded 4.0mg/l and 6.0mg/l for both seasons respectively.

## Plankton

Three classes of the microalgae group were represented in these rivers namely bacillariophyceae, Chlorophyceae and Cyanophyceae with the diatoms dominating the phytoplankton spectrum with total number of 41 taxa belonging to 16 genera out of the 59 taxa identified and most of it recorded in the wet season (Table 1). Green algae recorded 9 genera spread across the order zygnematales (*Spirogyra*) and desmidiiales (*Cosmarium*) while blue green was represented by 3 genera; *Anabaena*, *Oscillatoria* and *Spirulina*. Higher phytoplankton individuals (cells/ml) was recorded in Okpo River (Figure 1). The zooplankton were represented by three phyla, Cladocera, Copepoda and rotifera with *Bosmina longirostris* recording the highest number of species (17cells/ml) at Okpo River in the wet months with rotifers being the frequently encountered species (Table 2). Species evenness (j) and Shannon-Weiner ( $H^1$ ) indices followed almost the same pattern for phytoplankton as the highest and lowest values (1.38; 0.38) for Shannon-Weiner coincided with the highest and lowest values (0.50; 0.14) for species evenness and these values were recorded in wet and dry seasons respectively at Okpo river (Figure 2). For zooplankton, highest Shannon-Weiner value (1.12) and species evenness (0.99) were recorded at Okpo River while the Margalef index recorded the highest value (1.12) at Idebu River during the dry season (Figure 3).

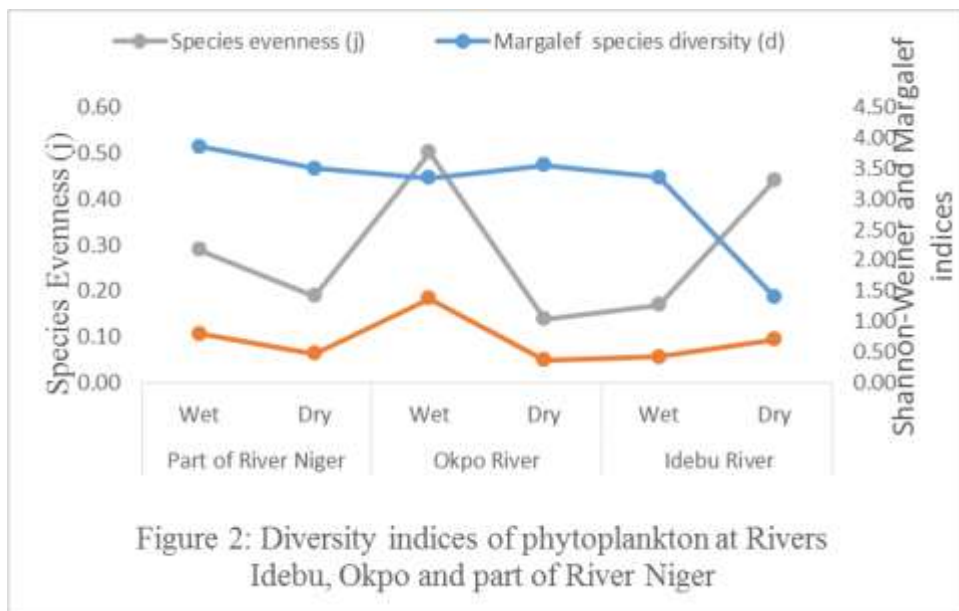
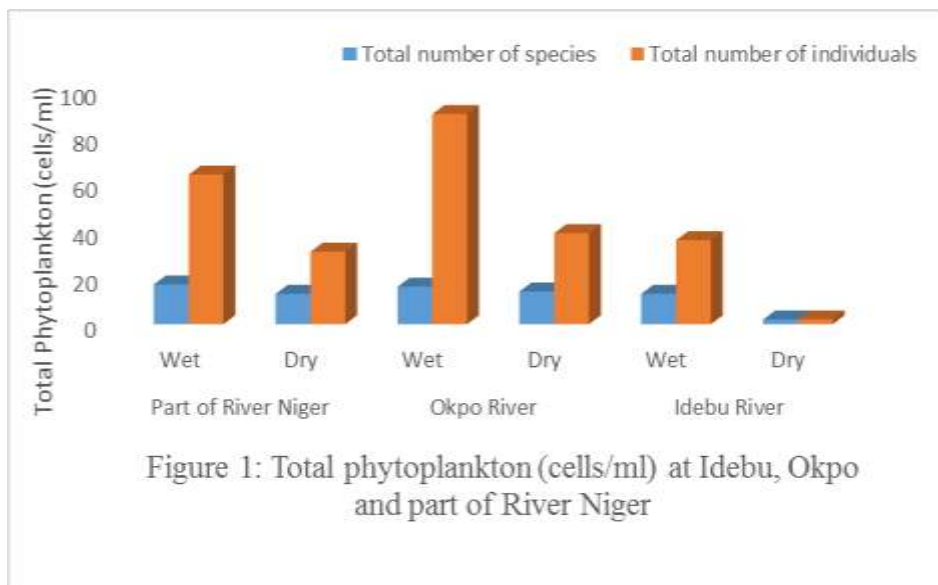
| <b>Table 1: Phytoplankton composition of Idebu, Okpo and Lower parts of River Niger (cells/ml).</b> |                                  |            |                   |            |                    |            |
|-----------------------------------------------------------------------------------------------------|----------------------------------|------------|-------------------|------------|--------------------|------------|
|                                                                                                     | <b>Lower Part of River Niger</b> |            | <b>Okpo River</b> |            | <b>Idebu River</b> |            |
| <b>CLASS:<br/>BACILLARIOPHYCEAE</b>                                                                 | <b>Wet</b>                       | <b>Dry</b> | <b>Wet</b>        | <b>Dry</b> | <b>Wet</b>         | <b>Dry</b> |
| <i>Achnanthes</i> sp (pennate)                                                                      |                                  |            |                   |            | 2                  |            |
| <i>Cymbella affinis</i> Kut                                                                         | –                                |            | 1                 |            |                    | –          |
| <i>C. ehrenbergii</i> Kutz                                                                          | 1                                |            | –                 |            |                    | –          |
| <i>C. gracilis</i> (Rabh.) Cleve                                                                    | –                                |            | –                 |            |                    | 3          |
| <i>C. minuta</i> Hilse                                                                              | –                                |            | 5                 |            |                    | 6          |
| <i>C. obscura</i> Krasske                                                                           | 10                               |            | –                 |            |                    | –          |
| <i>C. prostrata</i> (Berkeley) Cleve                                                                | 1                                |            | –                 |            |                    | –          |
| <i>C. silesiaca</i> Bleisch                                                                         | 2                                |            | –                 |            |                    | 2          |
| <i>Navicula bergenenis</i> Hoh                                                                      |                                  |            |                   | 3          |                    |            |
| <i>N. decussis</i> Oestrup                                                                          |                                  | 1          |                   |            |                    |            |
| <i>N. exigua</i> (Greg.) O. Muller                                                                  |                                  | 1          |                   |            | 2                  |            |
| <i>N. laevisissima</i> Kutzing                                                                      |                                  | 1          |                   |            |                    |            |
| <i>Navicula</i> sp.                                                                                 |                                  |            |                   | 6          | 5                  |            |
| <i>Surirella elegans</i> Ehrenberg                                                                  |                                  | 1          |                   | 1          |                    |            |
| <i>Surirella</i> sp.                                                                                |                                  |            |                   | 4          | 1                  |            |
| <i>Amphora aequalis</i> Krammer                                                                     |                                  |            |                   | 5          |                    |            |
| <i>A. coffeaeformis</i> (Agardh) Kutzing                                                            | 1                                |            | 12                |            | 5                  | –          |
| <i>A. commutata</i> Grunow                                                                          |                                  |            |                   | 2          |                    |            |
| <i>A. holsatica</i> Hustedt                                                                         |                                  |            |                   | 3          |                    |            |
| <i>A. normanni</i> Rabenhorst                                                                       | 2                                |            | –                 |            |                    | –          |
| <i>A. ovalis</i> Kutz                                                                               |                                  | 4          |                   |            |                    |            |
| <i>A. veneta</i> Kutzing                                                                            | –                                |            | 1                 | 2          |                    | 2          |
| <i>Amphora</i> sp                                                                                   |                                  |            |                   | 2          |                    |            |
| <i>Anomoeoneis</i> sp                                                                               |                                  |            |                   | 2          |                    |            |
| <i>Caloneis ventricosa</i> (Ehr.) Meister                                                           | 1                                |            | 1                 |            |                    | –          |
| <i>Cocconeis scutellum</i> Ehr.                                                                     | –                                |            | 1                 |            |                    | 4          |
| <i>Cocconeis</i> sp.                                                                                | 2                                |            | –                 |            |                    | –          |

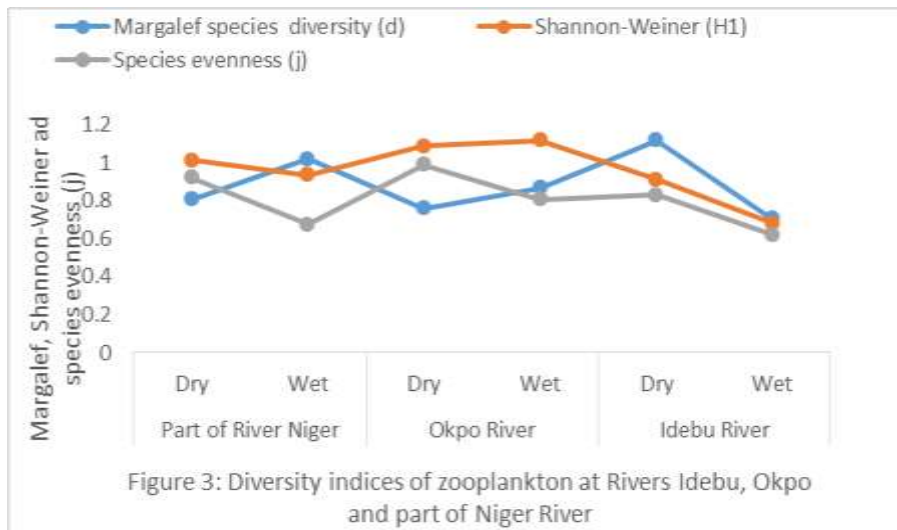
|                                                    |    |   |    |   |   |   |
|----------------------------------------------------|----|---|----|---|---|---|
| <i>Cyclotella meneghiniana</i> Kutz                | 2  |   | –  |   |   | – |
| <i>Diatoma</i> sp.                                 | –  |   | 1  |   |   | – |
| <i>Gonatozygon kinahanii</i> (Arch.)<br>Rabenhorst |    | 5 |    |   |   |   |
| <i>Gyrosigma scalproides</i><br>(Rabenhorst) Cleve |    |   |    |   | 1 |   |
| <i>Mastogloia braunii</i> Grunow                   |    | 1 |    | 2 | 3 |   |
| <i>M. smithii</i> Thwaites                         |    |   |    |   | 2 |   |
| <i>Neidium ampliatus</i> (Ehrenberg)<br>Krammer    |    |   |    | 3 |   |   |
| <i>N. minutissimum</i> Krasske                     |    | 2 |    |   |   |   |
| <i>N. septentrionale</i> Cleve – Euler             |    | 7 |    | 1 |   |   |
| <i>Nitzschia kutzingiana</i> Hilse                 |    |   |    |   | 2 |   |
| <i>P. gibba</i> Ehrenberg                          |    | 1 |    |   |   |   |
| <i>P. interrupta</i> W. Smith                      |    | 4 |    |   |   |   |
| <i>P. lundii</i> Hustedt                           |    |   |    |   | 4 |   |
| <i>P. maior</i> (Kutzing) Rabenhorst               |    | 1 |    |   |   |   |
| <b>CLASS: CHLOROPHYCEAE</b>                        |    |   |    |   |   |   |
| <i>Ankistrodesmus</i> sp.                          | –  |   | 1  |   |   | – |
| <i>Chlorella vulgaris</i> Beyerinck                | –  |   | 2  |   |   | – |
| <i>C. ellipsoidea</i> Gerneck                      | –  |   | 2  |   |   | – |
| <i>Closterium archerianum</i> Cleve                | 3  |   | –  |   |   | – |
| <i>C. ehrenbergii</i> Menegh                       | –  |   | 1  |   |   | – |
| <i>C. moniliferum</i> Ehrenb.                      | 1  | 2 | 3  |   | 3 | – |
| <i>C. pseudodiana</i> Roy                          | 2  |   | –  |   |   | – |
| <i>Closterium</i> sp.                              | 2  |   | –  |   |   | – |
| <i>Cosmarium botrytis</i> Meneg.                   | 1  |   | –  |   |   | – |
| <i>Cosmarium</i> sp.                               | –  |   | 55 | 3 |   | – |
| <i>Desmodesmus quadricauda</i><br>(Turp) Brebisson | 2  |   | –  |   |   | – |
| <i>Euasturm</i> sp.                                | 1  |   | –  |   |   | – |
| <i>Microspora</i> sp.                              | 30 |   | –  |   |   | – |
| <i>Scenedesmus bijuga</i> (Turp.)<br>Lagerherm     | –  |   | 1  |   |   | – |
| <i>Spirogyra</i> sp.                               | –  |   | 1  |   |   | – |

|                                |      |      |      |      |      |      |
|--------------------------------|------|------|------|------|------|------|
| <b>CLASS: CYANOPHYCEAE</b>     |      |      |      |      |      |      |
| <i>Anabaena</i> sp.            | –    |      | 2    |      |      | –    |
| <i>Oscillatoria</i> sp.        |      |      |      |      | 5    |      |
| <i>Spirulina</i> sp            |      |      |      |      | 1    |      |
| Total number of species        | 17   | 13   | 16   | 14   | 13   | 5    |
| Total number of individuals    | 64   | 31   | 90   | 39   | 36   | 17   |
| Margalef species diversity (d) | 3.85 | 3.49 | 3.33 | 3.55 | 3.35 | 1.41 |
| Shannon-Weiner ( $H^1$ )       | 0.81 | 0.48 | 1.38 | 0.38 | 0.43 | 0.71 |
| Species evenness (j)           | 0.29 | 0.19 | 0.5  | 0.14 | 0.17 | 0.44 |

Table 2: Zooplankton composition at Idebu, Okpo and Lower parts of River Niger

|                                  | Lower Part of River Niger |      | Okpo River |      | Idebu River |      |
|----------------------------------|---------------------------|------|------------|------|-------------|------|
|                                  | Dry                       | Wet  | Dry        | Wet  | Dry         | Wet  |
| <b>Cladocera</b>                 |                           |      |            |      |             |      |
| <i>Bosmina longirostris</i>      | –                         | 8    | 2          | 17   | 2           | 2    |
| <b>Copepoda</b>                  |                           |      |            |      |             |      |
| <i>Eucyclops agiloides</i>       | 2                         | 5    | 6          | 2    | 2           | 10   |
| <b>Rotifera</b>                  |                           |      |            |      |             |      |
| <i>Kellicortia longispina.</i>   | 4                         | 4    | 6          | 6    | 2           |      |
| <i>Branchionus diversicornis</i> | 6                         | 2    |            | 7    |             | 5    |
| Total number of species          | 3                         | 4    | 3          | 4    | 3           | 3    |
| Total number of individuals      | 12                        | 19   | 14         | 32   | 6           | 17   |
| Margalef species diversity (d)   | 0.80                      | 1.02 | 0.76       | 0.87 | 1.12        | 0.71 |
| Shannon-Weiner ( $H^1$ )         | 1.01                      | 0.94 | 1.09       | 1.12 | 0.91        | 0.68 |
| Species evenness (j)             | 0.92                      | 0.68 | 0.99       | 0.81 | 0.83        | 0.62 |



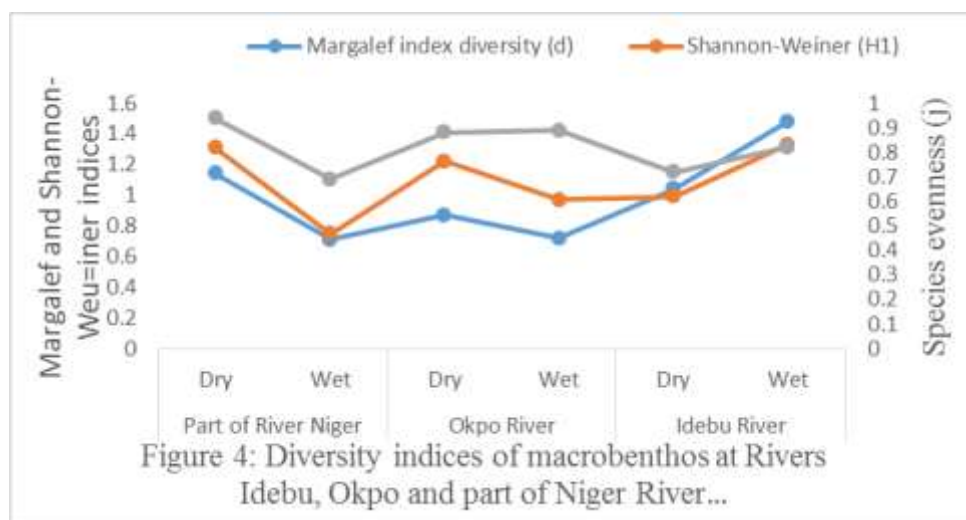


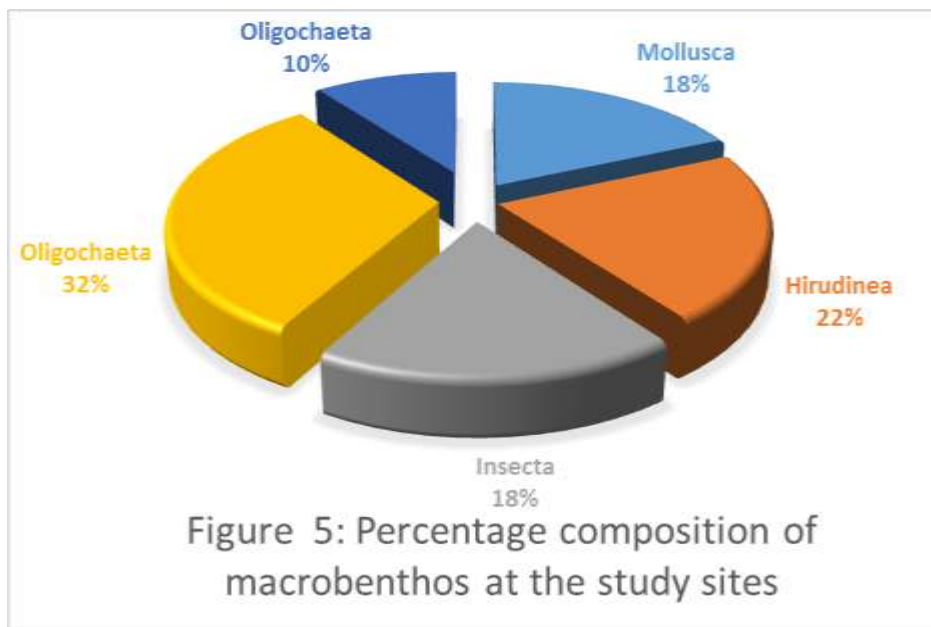
### Macrobenthos

The spatial distribution, species composition and diversity of macro-benthic fauna are presented in Table 3. Oligochaeta accounted for 37.27% of the total individuals with dominant taxon being *potamopyrgus* sp. followed by Hirudinea (19.09%); insect and mollusca had 16.36% each; and crustacean recorded 10.91%. Shannon-Weiner index ( $H^1$ ) and species richness (d) highest values (1.33; 1.48) were recorded at Idebu River while lowest values (0.75; 0.72) were observed at Okpo and a part of River Niger respectively (Figure 4). The part of River Niger during dry season recorded the highest (0.94) species evenness (j) value (Figure 4).

Table 3: The distribution, occurrence and diversity indices of macrobenthic invertebrate Community.

|                                | Lower Part of River Niger |      | Okpo River |      | Idebu River |      |
|--------------------------------|---------------------------|------|------------|------|-------------|------|
|                                | Dry                       | Wet  | Dry        | Wet  | Dry         | Wet  |
| <b>Mollusca</b>                |                           |      |            |      |             |      |
| <i>Potamopyrgus</i> sp.        |                           |      | 14         |      | 4           |      |
| <b>Hirudinea</b>               |                           |      |            |      |             |      |
| <i>Glossiphonia complanata</i> | 6                         | 1    |            | 2    | 8           | 4    |
| <b>Insecta</b>                 |                           |      |            |      |             |      |
| <i>Chrysops</i> sp.            | 4                         |      | 9          |      | 3           | 2    |
| <b>Crustacea</b>               |                           |      |            |      |             |      |
| <i>Palaemonetes</i> spp.       |                           |      | 2          | 6    |             | 4    |
| <b>Oligochaeta</b>             |                           |      |            |      |             |      |
| <i>Lumbriculus</i> sp.         |                           | 12   | 7          | 8    |             | 4    |
| <i>Tubifex</i> sp.             | 2                         | 4    |            |      | 3           | 1    |
| Number of species              | 3                         | 3    | 4          | 3    | 4           | 5    |
| Number of individual           | 12                        | 17   | 32         | 16   | 18          | 15   |
| Margalef index diversity (d)   | 1.14                      | 0.71 | 0.87       | 0.72 | 1.04        | 1.48 |
| Shannon-Weiner ( $H^1$ )       | 1.31                      | 0.75 | 1.22       | 0.97 | 0.99        | 1.33 |
| Species evenness (j)           | 0.94                      | 0.69 | 0.88       | 0.89 | 0.72        | 0.82 |





### Discussion

The relative abundance of diatoms in these rivers may be because they are able to grow at low light levels. The present observation that diatoms dominate the phytoplankton community confirms earlier reports made by Nwadiaro (1990) in the chanomi creek system of the Niger Delta; Chindah and Pudo (1991) in Bonny River; Erondy and Chindah (1991) in the new Calabar River, Niger Delta; Adesalu (2008) in Lekki lagoon; Adesalu and Nwankwo (2005, 2008) in Olero and Abule Eledu creek respectively; Adesalu *et al.* (2008, 2014) in Ogbe and Ipa-Itako creeks, and Nwankwo (1986, 1991) in the Lagoons of South western Nigeria. Many factors may have contributed to the dominance of pennate forms, which probably may be a confirmation of the shallow nature of these rivers. Differences in abundance of most species during the two sampling periods could be attributed to significant variations in the physico-chemical variables and nutrients levels within these periods. High number of individuals (cells/ml) recorded for these rivers in the wet season, could be because the rivers are seasonal and are almost getting dry during the dry season and getting

flooded in the wet season. The flooding will bring along with it flood water from the riparian vegetation and this probably explains the high cells/ml recorded during the wet season in this study. However, the number of zooplankton recorded in this study was scanty which may be as a result of the seasonal characteristics of these rivers as more, in terms of number, was recorded in the wet season. This is in conformity with Imoobe (2011) who stated that flooding during the rainy season may have contributed positively to zooplankton population growth as a result of species recruitment from other flooded waterbodies and the inflow of nutrients from the drainage basin that will trigger off increase in phytoplankton production and consequently zooplankton productivity. Higher number of zooplankton species richness recorded in the rainy season agreed with the findings of Imoobe (2011). According to Akinbuwa & Adeniyi, (1996); Imoobe and Akoma, (2009); and Imoobe (2011) the dominant status of rotifer species in rivers comparative to the cladocerans and copepods is characteristic of tropical lakes and rivers which this study conformed with. However, seasonal dynamics of zooplankton communities in the tropics has been attributed to a number of other factors such as the environmental characteristics of the water, predation, quality and quantity of edible algae and competition (Ovie and Adeniji 1994). It is obvious from this study that the environmental conditions especially rainfall were the main factors controlling the abundance of zooplankton. Macro-benthic communities play important roles in ecosystem processes such as recycling nutrients, detoxifying pollutants and secondary production (Snelgrove, 1997). However, the present study showed a dearth in number and composition of species. The family, lumbriculidae, has been reported to dominate aquatic benthic invertebrates' communities because they hardly show any habitat restriction (Victor and Ogbeibu, 1991) and are known to replace other invertebrate taxa in aquatic habitat perturbed by human activities. This probably indicates the extent of human activities around these rivers. The presence of oligochaetes supported Ogunwenmo and Kusemiju (2004) who reported that oligochaetes preferred polluted tropical waters and soft sediments rich in organic matter as observed in this study. Furthermore, the dominance by the deposit-feeding gastropod, *Potamopyrgus* sp. which are

common in all types of running water and are found in vast numbers on stones, vegetation or mud shows, a clearly opportunistic behavior (Bemvenuti *et al.*, 1992) which is also an indication of a stressed environment. However, the low number in the population of macro-benthic invertebrates recorded at the study stations could be due to the developmental rate of small macro-invertebrates, since most aquatic invertebrates are benthic only at larval stages while their adult lives are spent outside aquatic environments (Ibemenuga and Inyang, 2006).

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### **References**

- Abdullahi, B.A, and Indabawa I.I (2005). Ecology of Freshwater Phytoplankton of River Hadejia, Jigawa State, Nigeria. Biological and Environmental Sciences Journal for the Tropics, 1(2):141-146.
- Abubakar, M.M (2007). The phytoplankton community of a semi-and lake, Nguru Lake. Kano Journal for Science Education, 4(1): 16-21..
- Adesalu, T.A. (2008): Phytoplankton dynamics in relation to water quality indices in Lekki lagoon. Ph.D thesis.
- Adesalu, T.A, Nwankwo, D.I (2005). Studies on the phytoplankton of Olero creek and parts of Benin River, Nigeria. The Ekologia, 3 (2):21-30.
- Adesalu, T. A, and Nwankwo, D. I (2008). Effect of water quality indices on phytoplankton of a sluggish tidal creek in Lagos, Nigeria. Pakistan J. Biol. Sci. 11: 836-844.
- Adesalu, T.A, Abiola, T.O and Bofia, O.T (2008). Studies on the epiphytic algae associated with two floating aquatic macrophytes in a sluggish non-tidal polluted creek in Lagos, Nigeria. Asian J. Sci. Res. 1: 363-373.
- Adesalu, T.A., Adesanya, T., and Ugwuzor C (2014). Phytoplankton composition and water chemistry of a tidal creek (Ipa –Itako) part of Lagos Lagoon. Journal of Ecology and Natural Environment, 6 (11): 373-388.

- Akinbuwa, O., and Adeniyi, I.F (1996). Seasonal variation,distribution and interrelationships of rotifers in Opa Reservoir, Nigeria. *Afr. J. Ecol.* 34: 351-363, 1996.
- Ali, M.Z., Michael, A.A., and Yahuza, T. (2013). Physico-Chemical Characteristics and Phytoplankton Abundance of the Lower Niger River, Kogi State, Nigeria. *Journal of Environmental Science, Toxicology and Food Technology*, 2(4):31-37.
- Aneni, I.T, and Hassan, A.T. (2003). Effect of pollution on seasonal abundance of plankton in, Kudeti and Onireke Stream Ibadan, Nigeria. *The Zoologist* 2 (2): 76-83.
- A.P.H.A: (1998). Standard methods for the evaluation of water and waste water. Washington D.C. USA.
- Atobatele, O.E, Morenikeji, O.A., and Ugwumba, O.A. (2005) Spatial Variation in Physical and Chemical Parameters and Benthic Macro-invertebrate Fauna of River Ogunpa, Ibadan. *The Zoologist* 3: 58-67.
- Bellinger, E.G., and Siegel, D.C. (2010). *Freshwater algae identification and use as bioindicators*. John Wiley & Sons, Ltd. The atrium, Southern Gate, Chichester, West Sussex UK: 271.
- Biddanda, B, and Benner, R. (1997). Carbon nitrogen and carbohydrate fluxes during the production of particulate and dissolved organic matter by marine phytoplankton. *Limnology and Oceanography*, 42: 506-518.
- Bemvenuti, C.E, Cattaneo S., and Netto, S.A. (1992). Características estruturais da macro fauna bentônica emdois pontos da região estuarina da Lagoa dos Patos, RS., Brasil. *Atlântica*, 14: 5-28.
- Chindah, A.C., and Pudo, J. (1991). A preliminary checklist of algae found in plankton of bonny River in Niger Delta, Nigeria. *Fragm. Flor. Geobot.* 36: 112-125.
- Davis, C.C. (1955). *The marine and freshwater plankton*. Michigan State University Press, Michigan. 562pp.
- Desikachary, T.V. (1959). *Cyanophyta*, *Indian Council of Agric. Res.* New Delhi.
- Dimowo, B.O. (2013). Monthly spatial occurrence of phytoplankton and zooplankton in River Ogun, Abeokuta, Ogun State, Southwest

- Nigeria. International Journal of Fisheries and Aquaculture, 5(8):193-203.
- Edokpayi, C.A., and Lawal, M.O., Okwok, N.A. and Ogunwenmo, C.A. (2004). Physico-chemical and macrobenthic faunal characteristics of Kuramo water, Lagos, Southern Nigeria. Afr. J. Aquat. Sci., **29** (2): 235-241.
- Emmanuel, B.E., and Onyema, I.C (2007). The Plankton and fishes of a tropical creek in South-western Nigeria. Turk. J. Fish. Aquat. Sci. 7:105-113.
- Erondu, E.S., and Chindah, A.C. (1991). Physico-chemical and phytoplankton changes in a tidal freshwater station of the New Calabar River South Eastern Nigeria. Environ. Ecol. 9: 561-570.
- Goulden, M. and Few, R. (2011). Climate change, water and conflict in the Niger River Basin, International Alert and University of East Anglia.
- Hendey, N.I. (1964). An introductory account of algae of British Coastal waters. Her Majesty's Stationary Office.
- Hustedt, F. (1930-1937). Die. Kiselalgen 7. In Rabenhorst (Ed.) *Kryptogamen-flora von Deutschland Österreichs und der Schweiz*. A Kademische Vergesellschaftung M.L.H. Leipzig.
- Ibemenuga, KN. and Inyang, N. (2006). Macroinvertebrate Fauna of a Tropical Freshwater Stream in Nigeria. Animal Research International **3**(3): 553 – 561.
- Imoobe, T.O.T. (2011) Diversity and Seasonal Variation of Zooplankton in Okhuo River, a Tropical Forest River in Edo State, Nigeria. Centrepont Journal 17 (1): 37-51.
- Imoobe, T.O.T, Akoma, O.C. (2009). Spatial variations in the composition and abundance of zooplankton in the Bahir Dar Gulf of Lake Tana, Ethiopia. Afr. J. Ecol., 48: 72–77.
- Krammer, H. and Large-Bertalot, H. *Bacillariophyceae*. 1. Teil: Naviculaceae In: Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D., (Hrsgb.), *Süßwasserflora von Mitteleuropa*. Bd. 2 Fischer Verlag, Stuttgart. 876pp.
- Margalef, DR. (1951). Diversidad de especies en les comunideades natural *Public Institutte of Biologic Barcelona*, 9: 5 -27.

- Mathivanan, V. and Jayakumar, S. (1995). The studies on plankton fluctuation in a reservoir of Annamalainagar. Proceedings of the national symposium on recent trends in Indian wild life research, AVC college, Mayiladuthurai, Tamilnadu, India.
- Mohammad, M.A. and Saminu, M.Y. (2012) A water quality and phytoplankton of Salanta river Kano, Nigeria. Biological Science and Bioconservation, 4: 65-73.
- Needham, J.G., Needham P.R. (1962). A guide to the study of freshwater biology. Holden-Day Inc. San Francisco.
- Nwankwo, D.I. (1986). Phytoplankton of a sewage disposal site in Lagos Lagoon, Nigeria. Nigerian Journal of Biological Sciences. 1: 89-91.
- Nwankwo, D.I. (1991). A survey of the dinoflagellates of Nigeria I. Armoured dinoflagellates of Lagos Lagoon and associated tidal creeks. Nigerian Journal of Botany 4: 49-60.
- Ogbeibu, A.E. and Oribhabor, B.J. (2002). Ecological Impact of river impoundment using benthic macro-invertebrates as indicators. Water Research 36: 2427- 2436.
- Ogunwenmo, C.A. and Kusemiju, K. (2004). Annelids of a West African estuarine system. *J. Environ. Biol.*, 25 (2): 227-237.
- Olaniyan, C.I.O. (1969). The Seasonal variation in the hydrology and total plankton of the lagoon of South-Western Nigeria. Nig. J. Sci. 3: 101-119.
- Olaniyan, C.I.O. (1975). The seasonal variation in the hydrological and total plankton of the lagoons of South-western Nigeria. Nig. J. Mar. Res. 8: 36-59.
- Ovie, S.I. and Adeniji H.A (1994) Zooplankton and environmental characteristics of Shiroro Lake at the extremes of its hydrological cycle. *Hydrobiologia* 286: 175-182.
- Patrick, R. and Reimer C.W. (1966). The diatoms of the United States exclusive of Alaska and Hawaii. Vol. 1–Monographs. Academy of Natural Sciences, Philadelphia. 688pp.
- Peerapornpisal, Y.C., Chaiubol, J., Peckoh, H., Kraibut, M., Chorum, P., Wannathong, N., Ngearnpat, K., Jusakul, A., Thammathiwat, J., Chuanunta and Inthasotti, T. (2004) Monitoring of water quality

- in Ang Kaew reservoir of Chiang Mai University using phytoplankton as bioindicator from 1995-2002. *Chiang Mai journal of science* 31(1): 85-94.
- Pielou, E.C. (1975). The measurement of diversity to different types of biological collections. *J. Theor. Biol.* 13: 131-144.
- Quigley, M. (1977). *Invertebrates of Streams and Rivers-a key to identification*. Edward Arnold (Publishers) Ltd., London. 81pp.
- Shannon, C.E. and Weaver, W. (1963). *The mathematical theory of communication*. University of Illinois. Press, Urbana, 111-125p.
- Snelgrove, P. (1997). The importance of marine sediment biodiversity in ecosystem processes. *Ambio*, 26: 578-583.
- Townsend, C.R., Harper, J.D. and Begon, M. (2000). *Essentials of Ecology*. 3rd Edition. Blackwell Science London UK.
- Victor, R., Ogbeibu, A.E. (1991). Macroinvertebrate communities in the erosional biotope of an urban stream in Nigeria. *Tropical Zoology* 4: 1-12.
- Vollenweider, R. A. (1969). A manual of methods marine primary productivity in Aquatic environment. Handbook no.12, Blackwell Scientific Oxford.
- Whitford, L.A, Schumacher, G.J. (1973). *A manual of fresh water algae*. Sparks, North Carolina, USA.
- Wiafe, G., and Frid C.L.J. (2001). *Marine zooplankton of West Africa*. Darwin Initiative Report 5.120pp.
- Wimpenny, R.S. (1966). *The Plankton of the Sea*. Faber and Faber Limited, London, England.